

Original Article

Reduction of Purification Time of Polyspecific Equine F(ab)₂ Antivenom against Scorpion Envenomation

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ABSTRACT

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Background and Aims: In this study we improved the purification of immunoglobulins from equine antiserum raised against the venom of 6 types of scorpion species. Caprylic acid (octanoic acid), a fatty acid, was found to have no effect on the activity of the enzymes pepsin, which is used in antivenom purification to digest Fc fragment of immunoglobulins to obtain F(ab)₂.

Materials and Methods: A new method was developed for the production of F(ab)₂ antivenom whereby whole equine antiserum was mixed with equal amount of 0.15 M HCl and pH 3.4 with pepsin 660 mg/L of diluted antivenom and incubated for 4 h at 37°C. After digestion the pH were brought to 4.8 with sodium hydroxide solution (1.5 M) and then 1.5% caprylic acid and 10% ammonium sulfate was added and mixed for 60 minutes and passed through filter paper.

Results: Caprylic acid caused precipitation of albumin, and ammonium sulfate reduced turbidity of solution, resulting in a reduced protein load presented to the digestion enzymes culminating in substantial reductions in processing time.

Conclusions: The equine F(ab)₂ obtained using these novel caprylic acid methods were comparable in terms of yield, purity and specific activity to those obtained by multi-step and time consuming conventional salt fractionation with ammonium sulfate.

Introduction

Antivenoms against snakes and scorpions bites have been produced worldwide for about a century and in Iran these antivenoms were produced for about 70 years ago. However, there remain various aspects of salt fractionation in conventional production process that should be optimized. For example, the, processes of fractionation should be developed to improve the recovery, potency, tolerability and stability of the product [1]. In Razi Serum and Vaccine Research Institute (South-Western Iran), the therapeutic polyvalent equine immunoglobulin, following pepsin digestion of Fc fragment from IgG, F(ab')₂ fragments against scorpion venoms, has been fractionated by ammonium sulfate precipitation which is a time consuming and expensive method. Recently by caprylic acid precipitation at certain concentrations, albumin and other undesired impurities are precipitated and filtered by filter paper and discarded. Filtrate containing antibody F(ab')₂ fraction, can be concentrated by ultrafiltration or centrifugation or precipitated with ammonium sulfate and the precipitate is dialyzed against distilled water [2]. It has been observed that the protein aggregates formed and present in the antivenom product purified by conventional methods may be responsible for the adverse reactions observed in antivenom therapy [3]. Caprylic acid has been used to fractionate whole IgG [4] and also in the production of F(ab')₂ and Fab from ovine and equine sources [5]. It was first used to fractionate antivenom F(ab')₂ by Dos Santos et

al. [6]. Caprylic acid, at about 5% (v/v), precipitates non-Immunoglobulin proteins and leaves the antibody in the supernatant. Therefore it is not necessary to re-dissolve the antibody repeatedly performed in older method, a process which also decreases antibody recovery. Another benefit of the caprylic acid fractionation is that it is likely to inactivate lipid enveloped viruses potentially transmissible from horse to human and remove pyrogenic factors [7]. Moreover, this process is superior in terms of higher yield, less production time and less protein aggregation when compared with ammonium sulfate precipitation method [4]. The antivenom plasma in the presence of high concentration of caprylic acid becomes quite turbid [8] and removal of protein precipitate by filtration leaves a turbid filtrate. Sequential precipitation of F(ab')₂ fragment present in turbid filtrate with ammonium sulfate has been shown to be highly effective in the laboratory and production scale purification of antibody from various animal plasma fluids [9-11]. Undesired plasma proteins were first precipitated by caprylic acid and the precipitate was removed by filtration or centrifugation after removal of impurities. The supernatant containing antibody was then precipitated by ammonium sulfate and dialyzed against distilled water. This simple and fast process could result in antibody with purity comparable to that achieved by other techniques like ion-exchange chromatography [12].

It is interesting, therefore, to investigate the use of caprylic acid and ammonium sulfate in combination to fractionate equine antivenom $F(ab')_2$ [13] and see whether any improvement in terms of turbidity, yield, purity and the absence of protein aggregates could be achieved by the conventional method using ammonium sulfate. Since antivenom is mainly used in developing countries where cost of production is an important factor, the manufacturing process should be simple with minimal steps and be economical as well [14]. Therefore, in this study caprylic acid and ammonium sulfate were studied at concentrations that precipitated only non-antibody plasma proteins for the fractionation of horse $F(ab')_2$ antivenom. In order to simplify and minimize the manipulation steps involved in the fractionation, caprylic acid and ammonium sulfate were used sequentially without intermediate separation of the precipitate.

Materials and Methods

Chemicals

Lyophilized scorpion venoms of 6 species (*Hemiscorpius lepturus* belonging to *Hemiscorpionidae* family, *Androctonus crassicauda*, *Mesobuthus eupeus*, *Hottentuta saulcyi*, *Hottentuta schach*, *Odonthobuthus doriae* belonging to *Buthidae* family) produced in Razi Serum and Research Institute, Ahvaz branch, Iran were used for hyper-immunization of 20 female horses. Plasma was separated from whole blood by sedimentation using sodium citrate solution as anticoagulant with added preservative [15].

Chemicals and biochemicals were of reagent grade and were from Sigma-Aldrich (St-Louis, MO, USA) or Merck (Germany).

Conditions for pepsin digestion of horse plasma

Pepsin digestion of horse polyvalent plasma was carried out at 37°C, for 4 hours using pepsin (EC. 3.4.23.1) to plasma volume in the ratio of 660 mg per liter of initial plasma pH 3.5. The digestion was stopped by adding 1 M NaOH to bring the pH to 6.0.

Fractional precipitation of horse anti-scorpion $F(ab')_2$ by caprylic acid and ammonium sulfate

Using a preparative format, all manipulations were carried out at room temperature of about 26-27°C. Pepsin digested plasma (100 mL and pH 4.8) was divided into 6 parts. This study involved the fractionation of equine scorpion antivenom $F(ab')_2$ by combined stepwise ammonium sulfate and caprylic acid precipitation without intermediate separation of precipitate. Six conditions with combinations of ammonium sulfate (0-20% saturation) and caprylic acid (0-3.5% v/v), were tested. ammonium sulfate significantly reduced the turbidity raised by caprylic acid. High specific antibody activity was observed in the area containing 2-3.5% caprylic acid and 0-20% saturation with ammonium sulfate. Out of these results, 6 precipitation conditions were selected for detailed quantitative studies (Table 1). As seen in table 1, some combinations, like one with 1.5% caprylic acid and 15% ammonium sulfate and another with 3.5% caprylic acid and 0% ammonium sulfate, gave the highest fold purification (1.71 and

1.69) with antibody recoveries at 72.79% and 77.28%, respectively.

The combinations of caprylic acid and ammonium sulfate offered a benefit over caprylic acid alone in reduction of turbidity and in promotion of purity but not the recovery of antibody. The conditions giving more favorable overall results were with 3.5% caprylic acid alone and another with a combination of 1.5% caprylic acid and 15% ammonium sulfate. These preparations of F(ab')₂ were homogeneous and without protein aggregate.

Ammonium sulfate at various concentrations was added slowly to each row with vigorous shaking for 60 min. The final concentration of ammonium sulfate used in this experiment was 0-20% saturation. The ammonium sulfate treating digested plasma in each beaker was adjusted to pH 4.8 with 1.5 M HCl. caprylic acid was added slowly to each container to leave final concentrations of 0, 1.5, 3.5 % (v/v) with vigorous shaking for 60 min. A condition containing digested plasma with 0% ammonium sulfate and 3.5 caprylic acid was used as the standard reference. The turbidity of each container was measured. Turbidity was again measured after the plate was kept overnight at 4°C. The contents of all the containers were transferred onto the filter papers. The filtrates were collected in the clean containers. The protein concentration and antibody activity in the filtrates were determined.

Specific antibody

The enzyme-linked immunosorbent assay (ELISA) antibody titer of F(ab')₂ of each precipitation condition was determined by a

competitive ELISA. The procedure was modified as described by Morais and Massaldi [16] and Rial et al. [17]. Preparation of polyvinyl microtiter plate was according to the reported procedure [18]. Appropriate dilutions with different precipitation conditions were described [19]. ELISA plate was made according to the reported methods [20]. The units of activity in the microplate precipitation experiments were fixed with the sample dilution at 1:80 and absorbance of the sample was compared to that of the standard [21]. The unit of activity of the test tube precipitation experiment was defined as the sample dilution which had an absorbance equal to half the absorbance of conjugate alone (50% competition) [22]. The antibody unit was calculated using Prism 5, Graph Pad software program (trial version) [23].

Statistical analysis

All the tests were performed in five replications and were presented mean±standard deviation. Data were statistically analyzed using analysis of variance (ANOVA) and Duncan's multiple range test by means of SPSS (standard version 19.0 SPSS Inc, Chicago, IL, USA). There was a significant difference in the value of $p < 0.05$.

Results

From the sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) of the digested plasma protein profiles, IgG digested to F(ab')₂ was completely digested at 60 min, while albumin was completely digested at much shorter period in less than 15 min. Fractionation of pepsin-digested horse plasma

using ammonium sulfate and caprylic acid precipitation under a 1000 mL format fractionation of horse plasma was performed.

Turbidity of the ammonium sulfate and caprylic acid precipitated pepsin-digested plasma: The turbidity of the precipitation conditions is shown in Table 1. In the absence of caprylic acid, an increase in ammonium sulfate concentration did not noticeably affect the turbidity. However, in the absence of ammonium sulfate, the turbidity of the suspensions sharply increased when caprylic acid was added, and at 3.5% (v/v) caprylic acid, the highest turbidity was observed.

Protein concentration of the filtrates of ammonium sulfate and caprylic acid precipitated pepsin-digested plasma: Protein concentration of the filtrates of ammonium sulfate and caprylic acid precipitated pepsin-digested plasma is shown in Table 1. At 10% saturated ammonium sulfate, protein concentration in the filtrate decreased. When caprylic acid was used, it sharply decreased the protein concentration in the filtrate; the combination of ammonium sulfate and caprylic acid resulted in more effective protein precipitation, especially at higher reagent concentrations. While the digested plasma control contained the highest protein concentration (52.25 mg/mL), the filtrate of 0% saturated ammonium sulfate and 3.5% (v/v) caprylic acid treated plasma contained the lowest protein concentration (24.86 mg/mL), indicating that 66% of the digested protein had been precipitated.

ELISA antibody titer of the filtrate of the 10% ammonium sulfate saturation and 1.5%

caprylic acid precipitated pepsin-digested plasma of all the filtrates was the highest and the digested plasma control without any addition of caprylic acid and ammonium sulfate showed the highest antibody titer. This gradually decreased when ammonium sulfate and caprylic acid were added (Table 1). At 10% saturated ammonium sulfate and 1.5% (v/v) caprylic acid, the ELISA antibody titer in the filtrate was lowest, as compared to that in the digested plasma control. Specific antibody activity of the filtrate of ammonium sulfate and caprylic acid precipitated pepsin-digested plasma: The specific activity was lowest in the digested plasma control but was higher when caprylic acid and ammonium sulfate were added to selectively precipitate non-antibody proteins. At 10% saturated ammonium sulfate and 1.5% (v/v) caprylic acid, the highest specific activity, i.e, the highest degree of purification of F(ab')₂ was achieved.

Fractionation of pepsin-digested horse antivenom plasma using combination of ammonium sulfate and caprylic acid precipitation as studied in tubes: Six precipitation conditions which gave relatively low turbidity and high ELISA antibody titer and specific activities in the microplate experiment were selected (Table 1) for detailed quantitative studies. The turbidity (% solid) of the reaction mixtures was similar to that observed in the microplate experiment. While an increase in caprylic acid concentration resulted in increase in turbidity, ammonium sulfate was shown to decrease the turbidity caused by caprylic acid. The lowest turbidity (0.4158%) was observed

in 20% ammonium sulfate and 1.5% caprylic acid precipitation.

The protein concentrations in the filtrates decreased significantly when precipitation was carried out at higher concentrations of caprylic acid and the presence of ammonium sulfate increased the amount of protein precipitate (Fig. 1 and Fig. 2). When the precipitation was left overnight, slightly lower protein concentration was found in the filtrates, indicating that more protein was precipitated antibody recovery. The presence of ammonium sulfate further increased the specific activity and decreased the antibody recovery. The highest fold-purification of 1.69 and 1.71 were observed at 3.5% caprylic acid and at 1.5% caprylic acid with 15% ammonium sulfate, respectively, but with antibody

recoveries of only 77.28% and 72.79%. Highest antibody recoveries of 81.65% and 79.45% were observed at 10% ammonium sulfate and 1.5% caprylic acid and 20% ammonium sulfate and 1.5% caprylic acid, respectively, with 1.60 and 1.61 fold purification (Table 1). The lowest protein content was observed in the condition of 20% saturation with ammonium sulfate and 2% caprylic acid. The turbidity of the mixtures of these precipitation conditions were in the medium range of about 0.41%-0.49% which was considerably lower than that of 3.5% caprylic acid alone (0.6459%). Comparison of SDS-Page of products purified with ammonium sulfate (old method) and those purified with caprylic acid are shown in Fig. 3.

Table 1. Quantitative estimation of volume, protein, antibody specific activity, antibody recovery and turbidity of fractions obtained from precipitation at various ammonium sulfate and caprylic acid concentrations

Condition		Volume of Filtrate	Protein of Filtrate	Total Protein	Antibody Recovery	specific Activity	Fold Purification	Turbidity
%AS	%CA	(mL)	(mg/mL)	(mg)	(%)	(unit/mg)		
0	0	870±5.61 ^e	52.14±0.47 ^d	45.16±1.09 ^d	100.00±0.00 ^f	305.19±0.04 ^a	1.00±0.00 ^a	0.0000±0.0000 ^a
0	3.5	774.6±5.54 ^c	24.528±0.95 ^b	19.00±0.86 ^b	77.28±1.10 ^c	526.93±12.80 ^c	1.69±0.02 ^d	0.6459±0.00273 ^f
10	1.5	796.2±1.64 ^d	27.764±0.49 ^c	22.41±0.65 ^c	81.65±0.85 ^e	484.28±8.23 ^b	1.60±0.008 ^c	0.4957±0.00213 ^d
15	1.5	725.4±2.70 ^b	25.258±0.39 ^b	18.3±0.31 ^b	72.79±0.84 ^b	537.89±20.70 ^d	1.71±.008 ^d	0.4545±0.00219 ^c
20	1.5	794.6±5.31 ^d	28.05±0.87 ^c	21.96±0.42 ^c	79.45±0.71 ^d	493.60±4.67 ^b	1.61±0.007 ^c	0.4158±0.00219 ^b
20	2.0	649±17.63 ^a	20.00±0.08 ^a	12.98±0.39 ^a	59.45±0.60 ^a	519.23±4.78 ^c	1.52±0.03 ^b	0.5819±0.01206 ^e

Values are expressed as mean±SD (n=5), and values in the same line with different superscripts (a-e) are differences significant at p<0.05 by Duncan test using SPSS. (All of reaction voloum:1000 mL). AS=Ammonium sulfate; CA= Caprylic acid



Fig. 1. SDS-PAGE of purified F(ab')₂ fractionated with ammonium sulfate and caprylic acid at various concentrations. The electrophoresis was carried out in 10% acrylamide under reducing condition.

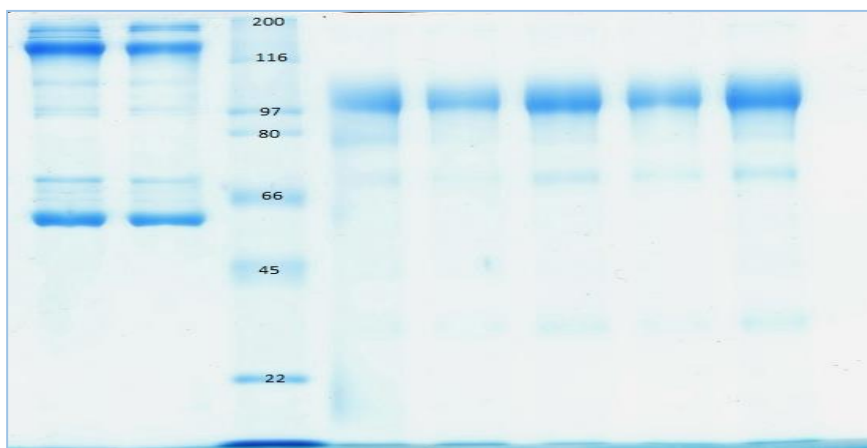


Fig. 2. SDS-PAGE of purified F(ab')₂ fractionated with ammonium sulfate and caprylic acid at various concentrations. The electrophoresis was carried out in 10% acrylamide under non-reducing condition.

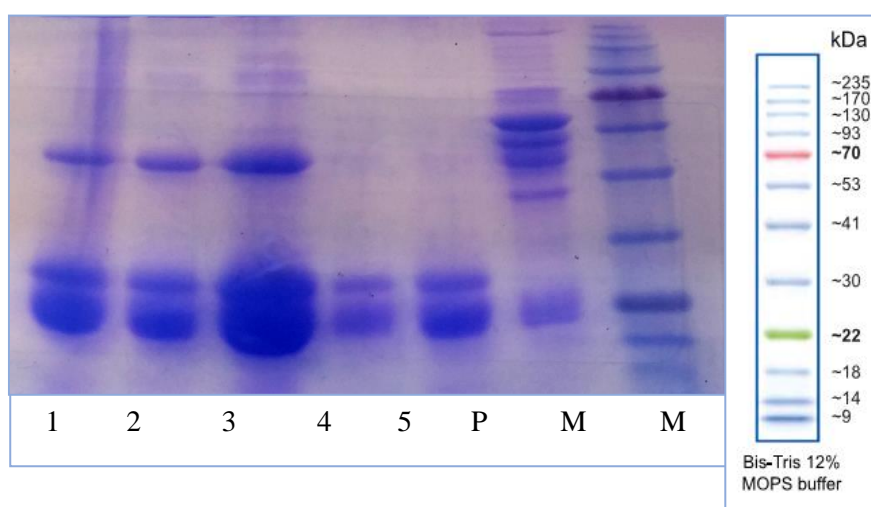


Fig. 3. Comparison of scorpion plasma purification. Lanes 1, 2 and 3: Experimental purification with ammonium sulfate alone (old method), 4 and 5: Experimental purification with caprylic acid and ammonium sulphate, P: Plasma, M: Marker.

Discussion

Large scale production of therapeutic antivenom against scorpion sting generally includes pepsin digestion of equine hyperimmune plasma and then fractionation of the $F(ab')_2$ fragment with ammonium sulfate. [22]. Intravenous administration of antivenoms is connected with a high (10-76%) prevalence of negative effects [3]. Additional contaminants considered to result in negative effects consisting of serum proteins and other fragments or excessive molecular weight aggregates. Consequently, fractionation of antivenom antibody has aimed at clearing the Fc portion by pepsin digestion and also eliminating additional plasma protein impurities or their digestion products by numerous procedures [18,19]. Pepsin digestion has been utilized since 1939 to eliminate the greatly immunogenic Fc of heterologous antibody. Nevertheless, it is essential for the pepsin to be entirely inactivated or eliminated in subsequent fractionation procedures because the enzyme can influence the stability of the antivenom [14]. Ammonium sulfate is commonly used in large-scale fractional precipitation of IgG or $F(ab')_2$. The negative aspect of ammonium sulfate precipitation is usually that the $F(ab')_2$ product needs to be recovered and resolubilized. This method is hard to perform on a large scale under aseptic situations, and endproduct pollution is usually discovered. The procedure can also involve an important decrease of antibody activity [1]. Incorporation of caprylic acid with ammonium sulfate fractionation has the benefit of

precipitating numerous serum proteins or their fragments. Considerably, the high molecular weight aggregates were not seen in the $F(ab')_2$ end product purified by caprylic acid precipitation. The risk of viral contamination of biological products is a subject of great interest [7]. In this regard, the fractionation studied regarding pepsin digestion and also treatment with caprylic acid might inactivate several viruses because of the acidic situation and the detergent action of the organic acid [22]. Since the purification of antibody is generally performed by dialysis in cellulose bags, this procedure is time-consuming and can take several days. With regards to the above notes, the aim of the present study was to prepare equine antivenom using combination of caprylic acid and ammonium sulfate methods as a novel and effective combinative method in improving of $F(ab')_2$ antivenom.

Conclusion

The present study made an attempt to purify equine antivenom using combination of caprylic acid and ammonium sulfate fractionation method in improving pepsin digested horse $F(ab')_2$ antivenom. The results of this study showed that using caprylic acid and then ammonium sulfate can be effective in removal of impurities and extraction of $F(ab')_2$ at high volumes in shorter period of time, which is an important factor in production of biological products. we used this method for purification of 50 liters of hyperimmune

plasma. The most important benefits of this method are 1) cheaper production cost which is an important factor in poor developing countries, 2) The shorter purification time which is the most important factor in biological product purification, and 3) removal or inactivation of progeny and protein aggregation and viral. Our results was consistent with those of other researchers who

asserted this method as effective in antivenom large scale purification.

Conflict of Interest

The authors declare that they have no conflict of interest.

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References

- [1]. Gutiérrez JM, León G, Burnouf T. Antivenoms for the treatment of snakebite envenomings: the road ahead. *Biologicals* 2011; 39(3): 129-42.
- [2]. Redwan ER. Comparison between therapeutic antitoxin F(ab)₂ fractionated with ammonium sulfate and caprylic acid. *J Immunoassay Immunochem.* 2006; 27(4): 319-29.
- [3]. Otero R, Gutiérrez JM, Rojas G, Núñez V, Díaz A, Miranda E, et al. A randomized blinded clinical trial of two antivenoms, prepared by caprylic acid or ammonium sulphate fractionation of IgG, in Bothrops and Porthidium snake bites in Colombia: correlation between safety and biochemical characteristics of antivenoms. *Toxicon.* 1999; 37(6): 895-908.
- [4]. Rojas G, Jiménez J, Gutiérrez J. Caprylic acid fractionation of hyperimmune horse plasma: description of a simple procedure for antivenom production. *Toxicon.* 1994; 32(3): 351-63.
- [5]. Krifi M, Ayeb MEL, Dellagi K, Venom J. The improvement and standardization of antivenom production in developing countries: comparing antivenom quality, therapeutical efficiency, and cost. *J Venom Anim Toxins.* 1999; 5(1-2): 128-41.
- [6]. Dos Santos M, Lima MDI, Furtado G, Colletto G, Kipnis T, Da Silva WD. Purification of F(ab)₂ anti-snake venom by caprylic acid: a fast method for obtaining IgG fragments with high neutralization activity, purity and yield. *Toxicon.* 1989; 27(3): 297-303.
- [7]. Burnouf T, Griffiths E, Padilla A, Seddik S, Stephano MA, Gutiérrez JM. Assessment of the viral safety of antivenoms fractionated from equine plasma. *Biologicals* 2004; 32(3): 115-28.
- [8]. Fernande AS, Kaundinya JO, Daftary G, Saxena L, Banerjee S, Pattnaik P. Chromatographic purification of equine immunoglobulin GF(ab)₂ from plasma. *J Chromatogr B.* 2008; 876(1): 109-115.
- [9]. Perosa F, Carbone R, Ferrone S, Dammacco F. Purification of human immunoglobulins by sequential precipitation with caprylic acid and ammonium sulphate. *J Immunol Methods* 1990; 128(1): 9-16.
- [10]. Reik LM, Maines SL, Ryan DE, Levin W, Bandiera S, Thomas PE. A simple, non-chromatographic purification procedure for monoclonal antibodies Isolation of monoclonal antibodies against cytochrome P450 isozymes. *J Immunol Methods* 1987; 100(1-2): 123-30.
- [11]. Temponi M, Kageshita T, Perosa F, Ono R, Okada H, Ferrone S. Purification of murine IgG monoclonal antibodies by precipitation with caprylic acid: comparison with other methods of purification. *Hybridoma* 1989; 8(1): 85-95.
- [12]. McKinney MM, Parkinson A. A simple, non-chromatographic procedure to purify immunoglobulins from serum and ascites fluid. *J Immunol Methods* 1987; 96(2): 271-80.
- [13]. Saetang T, Suttijitpaisal P, Ratanabanangkoon K, Nat J. Preparations of toxic components from *Naja kaouthia* venom by selective heat denaturation. *Toxins* 1998; 7(1): 37-44.
- [14]. Laemmli UK. leavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* 1970; 227(5259): 680-85.
- [15]. Slovis N, Murray G. How to approach whole blood transfusions in horses. *AAEP Proceedings* 2001; 47(8): 266-69.
- [16]. Morais V, Massaldi H. Effect of pepsin digestion on the antivenom activity of equine immunoglobulins. *Toxicon.* 2005; 46(8): 876-82.
- [17]. Rial A, Morais V, Rossi S, Massaldi H. A new ELISA for determination of potency in snake antivenoms. *Toxicon.* 2006; 48(4): 462-66.

- [18]. Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. Protein measurement with the Folin phenol reagent. *J Biol Chem.* 1951; 193(1): 265-75.
- [19]. Baldwin RL. How Hofmeister ion interactions affect protein stability. *Biophys J.* 1996; 71(4): 2056-63.
- [20]. Bernard N, Jolival C, Schwartzenruber J. Protein precipitation by caprylic acid: equilibrium composition data. *Biotechnol Bioeng.* 1996; 49(4): 405-11.
- [21]. Kukongviriyapan V, Poopyruchpong N, Ratanabanangkoon K. Some parameters of affinity chromatography in the purification of antibody against *Naja naja siamensis* toxin 3. *J immunol methods.* 1982; 49(1): 97-104.
- [22]. Raweerith R, Ratanabanangkoon K. Fractionation of equine antivenom using caprylic acid precipitation in combination with cationic ion-exchange chromatography. *J Immunol Methods* 2003; 282(1-2): 63-72.
- [23]. World Health Organization. WHO guidelines for the production, control and regulation of snake antivenom immunoglobulins. Geneva: WHO. 2010; p.134.