

Original Article

Plasma Iron Indices in Pregnant Women Referred to the University of Abuja Teaching Hospital, Nigeria

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ABSTRACT

Article history

Received: 26 Apr 2020

Accepted: 16 Aug 2020

Available online: 27 Aug 2020

Keywords

Anemia

Iron metabolism

Nigeria

Nutritional disorder

Pregnancy

Background and Aims: Iron Deficiency (ID) is a common obstetric problem and nutritional disorder that occurs mostly in developing countries. Hence, nutritional studies are required every few years to determine the necessary healthcare interventions for pregnant women. This cross-sectional study evaluated the plasma iron, ferritin, transferrin, total iron-binding capacity (TIBC), and unsaturated iron-binding capacity (UIBC) levels of pregnant women referred to the University of Abuja Teaching Hospital, Gwagwalada, Abuja, Nigeria.

Materials and Methods: Blood samples were collected from 58 non-pregnant women, 22 pregnant women in the first trimester, 52 in second, and 44 in the third trimester. The plasma ferritin concentration of these samples was measured by chemiluminescence assay, while the plasma transferrin, TIBC, and iron concentrations were measured by enzyme-linked immunosorbent assay. Plasma ferritin levels $<15\mu\text{g/L}$ were considered ID.

Results: The overall prevalence of ID in pregnant women was 33.1%. However, the prevalence of ID was 29.3%, 22.7%, 34.6%, and 36.4% among non-pregnant women, women in the first trimester, second and third trimester, respectively. The mean \pm SEM iron levels were significantly higher among pregnant women compared to non-pregnant women ($p=0.004$). There was no significant difference in the mean \pm SEM of plasma ferritin, transferrin and TIBC concentrations between pregnant and non-pregnant women ($p>0.05$). Nevertheless, the mean \pm SEM plasma iron, ferritin, and UIBC significantly differed across the gestational ages of pregnant women ($p<0.05$).

Conclusions: This study revealed a high prevalence of ID during pregnancy, which increases with the trimester of affected women. Healthcare interventional measures that can address ID are recommended.

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Introduction

With about one-gram requirement for the expansion of maternal red cell mass, placental, and fetal development, iron becomes essential for a healthy pregnancy [1]. Increasingly, severe iron deficiency and iron-deficiency anemia in pregnancy have been linked to higher chances of adverse maternal and fetal outcomes, including maternal mortality, perinatal death, preterm birth, low birth weight, impaired immune function, and long-term cognitive defects in newborns and infants [2]. In a bid to forestall these adverse outcomes, the World Health Organization (WHO) has recommended daily iron supplementation for all women during pregnancy [3]. On the other hand, the implications of high maternal iron levels in pregnancy are still poorly understood [4].

Iron in excess can lead to generating harmful reactive oxygen species (ROS) that are deleterious to proteins, lipids, and nucleic acids [5], as well as increased susceptibility to infection [6]. Maternal plasma ferritin, which is a marker of body iron stores, is linked to adverse pregnancy outcomes in a U-shaped manner [7]. Increased levels of plasma ferritin, however, are not only pointing to increased iron stores but additionally to inflammation. Besides, the exact contribution of excess iron is still needed to be elucidated. Despite its importance, efforts are still ongoing to accurately discern the regulation of maternal and fetal iron homeostasis in pregnancy, as well as the relative contribution of the maternal, placental, and fetal signals.

The cut-off marks for establishing anemia in pregnancy are hemoglobin concentration levels lower than 11.0 g/dL during the first and third trimester of gestation or lower than 10.5 g/dL in the second trimester of pregnancy [8]. The above slight differences in values stem from the plasma volume expansion of 40 to 50% exceeding the 20 to 25% increase in red cell mass, leading to physiological haemo-dilution [9]. This physiologic plasma expansion has been associated with favorable pregnancy outcomes [10]. The reason for the adoption of the 10.0 g/dL cut-off level in developing countries was because adverse fetomaternal outcomes are not usually found >10.0 g/dL hemoglobin level in these countries [8]. The estimated global prevalence of anemia in pregnancy is 41.8% [11]. With its high incidence, it becomes pertinent to assay the level of iron metabolism in pregnant women regularly. It means that when the mother becomes anemic, this transfer reduces, and the fetus will be at risk of deficiency [12].

At the apical surface of the syncytiotrophoblast, transferrin receptor 1 (TFR1) takes up iron-transferrin (Fe-Tf) from the maternal circulation [13]. The complex of Fe-Tf and TFR1 is endocytosed, and iron is released in the endosome and subsequently will be exported on the basal side of the placental syncytiotrophoblast through the iron transporter ferroportin (FPN) into the fetal circulation [5]. Both TFR1 and FPN are thought to be critical for iron transport across the placenta because the global deletion of either transporter leads to

embryonic death. About 75% of all cases of non-physiologic anemia in pregnancy roots in iron deficiency [2].

In the laboratory, complete cell blood count provides the basis for the diagnosis of iron deficiency anemia. Other useful tests include the determination of the levels of plasma iron, total iron-binding capacity, and transferrin [14]. Iron deficiency anemia (IDA) has increasingly been associated with high chances of preterm delivery, postpartum hemorrhage, low birth weight, and delayed psychomotor development in infancy [14-16]. On the other hand, other studies have revealed conflicting conclusions regarding specific outcome measures [17, 18]. Indeed, several studies across Nigeria have investigated IDA in pregnancy. However, little or none have comprehensively evaluated the iron metabolism indices. In this regard, there is every need to assess the current plasma iron metabolism indices as a measure of iron deficiency anemia among pregnant women attending the University of Abuja Teaching Hospital, Gwagwalada, Abuja, Nigeria. It will help to inform policies on preventive strategies and optimization of maternal and fetal outcomes during and after pregnancy.

Materials and Methods

The current study was a hospital-based cross-sectional study conducted on pregnant women who consented to participate in the study at the antenatal clinic at the University of Abuja Teaching Hospital (UATH), Gwagwalada, Abuja, Nigeria.

This study was carried out at UATH, which is a tertiary referral hospital situated in

Gwagwalada, a suburb from the Federal Capital Territory (FCT) of Nigeria. The University of Abuja Teaching Hospital has an average of 3,000 deliveries annually.

The minimum sample size for this study was determined from a previous cross-sectional study done by Ajepe et al. [19], which reported 12.3% iron deficiency among pregnant women in Nigeria. The calculated size at a 95% confidence interval was 170 individuals. However, a total of 176 subjects, which comprised of 118 pregnant women and 58 non-pregnant, were recruited.

The present study involved a total of one hundred and eighteen pregnant and fifty-eight non-pregnant women. Among the aged-matched pregnant women, 22 were on their first trimester, 52 and 44 were in their second and third trimester, respectively. They were selected based on having no history of surgical operation, malaria, HIV and significant inflammation. Besides, non-pregnant women were not on menstruation. All participants voluntarily consented to be enrolled in this study, and self-reported adherence with routine oral iron supplementation and completion of intermittent malarial prophylaxis at least for four weeks interval after the first trimester were considered as the inclusion criteria. In Nigeria, the routine iron supplementation program exists during antenatal clinic visits. However, some women (especially rural residents) do not adhere to their appropriate consumption due to ignorance and certain sociocultural beliefs. Ethical approval was obtained from the Human Research Ethical Committee (HREC) of the UATH. Written consent was also obtained

from all participants. This study was conducted following the standards of human experimentation described in the Helsinki Declaration of 1975 (as revised in 2000).

Because of the circadian fluctuation of iron, three millilitres (3mL) of whole fasting blood samples were collected from all participants and carefully dispensed into lithium heparin anticoagulant container. It was spun at 10,000 g for 5 minutes to ensure plasma separation, and the harvested plasma samples were used for the quantification of the analytes of interest. Samples were analyzed in batches within 1 hour of collection.

Laboratory analytical protocol

a. Quantification of plasma ferritin by sandwich Chemiluminescence Assay (CLIA). This study was assayed using Maglumi® (Snibe Co., Ltd, China), and Ferritin CLIA Kit (Catalog No. 130201001M). The assay was done based on the kits' instruction manual.

The anti-ferritin monoclonal antibody was used to label N-(4-amiobutyl)-N-ethylisoluminol (ABEI), and another monoclonal antibody was used to label fluorescein isothiocyanate (FITC). Samples, calibrators and controls were mixed thoroughly with FITC label and nanomagnetic microbeads in a cuvette incubated at 37°C, then cycle washing was carried out once, which was followed by the addition of ABEI label, incubated and washed for a second time. Subsequently, the starter reagents were added, and a flash chemiluminescent reaction was initiated. A photomultiplier measured the light signal as RLU within 3 seconds, and it was proportional to the concentration of ferritin present in samples.

b. Total iron-binding capacity (TIBC) and plasma iron assays were done using an Abcam® (UK) enzyme-linked immunosorbent assay (ELISA) Kit (Catalogue No. ab239715). It provided a simple, sensitive and high-throughput adaptable approach to quantify TIBC and plasma iron concentration. The assay was done, and the results were interpreted based on kits manufacturer's instructions.

c. The unsaturated iron-binding capacity (UIBC) was calculated by subtracting the TIBC concentration from plasma iron concentration.

d. The plasma transferrin concentration was determined by using Human Transferrin SimpleStep Abcam® (UK) ELISA Kit (Catalogue No. ab187391). The assay was completed, and the results were read based on the kit manufacturer's instructions.

Statistical analysis

Data generated were analyzed by Statistical Package for Social Science (SPSS version 22). Statistical differences between mean and standard deviation were determined using the student's t-test (for two groups) and ANOVA (for three groups or more). The association between iron deficiency (ID) and pregnancy status of participants was determined by univariate logistic regression. P values < 0.05 at 95% confidence interval were considered as statistically significant.

Results

The mean±SEM age (years) of these women were 25.53±0.70, 31.05±1.03, 30.9±0.61, 29.32±0.60, for the non-pregnant women, those in their first trimester, second and third trimester, respectively. The overall prevalence

of ID in pregnant women was 33.1%. The prevalence of ID was 29.3%, 22.7%, 34.6%, and 36.4% among non-pregnant women, women in their first trimester, second and third trimester, respectively (Table 1). There was no significant association between the prevalence of ID and the trimester of pregnant women ($p>0.05$).

The mean \pm SEM iron levels among pregnant women were 13.65 ± 0.29 $\mu\text{mol/L}$, which was significantly higher than that of non-pregnant women with 12.09 ± 0.48 $\mu\text{mol/L}$ ($p=0.004$) (Fig. 1). The mean \pm SEM ferritin level among the pregnant women was lower, 26.88 ± 1.99 $\mu\text{g/L}$ when compared to the non-pregnant women, 32.93 ± 3.81 $\mu\text{g/L}$. However, there was no significant difference in the plasma levels of ferritin between pregnant women compared to their non-pregnant counterparts ($p=0.123$) (Fig. 2).

There was a significantly lower mean \pm SEM unsaturated iron-binding capacity (UIBC) level in pregnant women, 53.87 ± 1.19 $\mu\text{mol/L}$ in comparison with the non-pregnant women, 65.55 ± 2.20 $\mu\text{mol/L}$ ($p<0.001$) (Fig. 1). The TIBC was lower, but it was not significantly different among pregnant women, 68.23 ± 3.53 $\mu\text{mol/L}$ as compared to non-pregnant women, 77.71 ± 2.61 $\mu\text{mol/L}$ ($p=0.079$) (Fig. 1). The mean \pm SEM transferrin level among the pregnant women (17.62 ± 0.69 $\mu\text{mol/L}$) was higher, but it was not significantly different compared to their non-pregnant counterparts (16.25 ± 0.90 $\mu\text{mol/L}$) ($p=0.239$) (Fig. 1).

After the categorization of the trimester of pregnant women, the mean \pm SEM plasma levels of iron, ferritin, and UIBC significantly

differed from one another ($p<0.05$). Nevertheless, there was no significant difference between the mean \pm SEM plasma levels of TIBC and transferrin of all the four categories of the participants ($p>0.05$) (Table 2). The mean \pm SEM plasma level (12.09 ± 0.48 $\mu\text{mol/L}$) of iron among the non-pregnant women was significantly lower than that of pregnant women in their second trimester, 13.92 ± 0.48 $\mu\text{mol/L}$ ($p=0.005$) and third trimester, 13.64 ± 0.42 $\mu\text{mol/L}$ ($p=0.021$). However, the mean \pm SEM plasma iron level among the non-pregnant women did not significantly differ from that of pregnant women in first trimester, 13.01 ± 0.65 $\mu\text{mol/L}$ ($p=0.270$) (Table 3).

The mean \pm SEM plasma ferritin level among the non-pregnant women, 32.93 ± 3.81 $\mu\text{g/L}$, was not significantly lower than that of pregnant women in their first trimester, 38.50 ± 5.50 $\mu\text{g/L}$ ($p=0.356$), but higher than those in the second trimester, 24.19 ± 2.52 $\mu\text{g/L}$ ($p=0.059$) and third trimester, 24.25 ± 3.29 $\mu\text{g/L}$ ($p=0.073$) (Table 3). The mean \pm SEM plasma UIBC level among the non-pregnant women was 65.54 ± 2.20 $\mu\text{mol/L}$, which was not significantly higher than those in the first trimester ($p=0.086$). However, it was significantly higher compared to those in the second trimester, 51.86 ± 1.66 $\mu\text{mol/L}$ ($p<0.001$) and those in the third trimester, 53.47 ± 2.15 $\mu\text{mol/L}$, respectively ($p=0.000$) (Table 3).

Table 1. The prevalence of iron deficiency by pregnancy status of participants

Pregnancy status	No. of participants	Iron Deficiency (%)	Odds Ratio (95% CI)	P-value
Non-pregnant	58	17 (29.3)	Referent	
First trimester	22	5 (22.7)	0.71 (0.23-2.23)	0.557
Second trimester	52	18 (34.6)	1.28 (0.57-2.85)	0.551
Third trimester	44	16 (36.4)	1.37 (0.59-3.18)	0.451

Significant association determined by univariate logistic regression

Table 2. Comparison of Iron Indices between non-pregnant and the various trimesters of pregnant women

Iron indices	Non-pregnant state (n=58)	First trimester (n=22)	Second trimester (n=52)	Third trimester (n=44)	F value	P-value
Iron ($\mu\text{mol/L}$)	12.09 \pm 0.48	13.01 \pm 0.65	13.92 \pm 0.48	13.64 \pm 0.42	3.206	0.025*
Ferritin ($\mu\text{g/L}$)	32.93 \pm 3.81	38.50 \pm 5.50	24.19 \pm 2.52	24.25 \pm 3.29	2.928	0.035*
Unsaturated iron-binding Capacity ($\mu\text{mol/L}$)	65.55 \pm 2.20	59.41 \pm 2.37	51.86 \pm 1.66	53.47 \pm 2.15	10.227	0.000*
Total iron binding Capacity ($\mu\text{mol/L}$)	77.71 \pm 2.61	71.71 \pm 2.55	63.89 \pm 1.90	71.62 \pm 9.14	1.556	0.202
Transferrin ($\mu\text{mol/L}$)	16.25 \pm 0.90	17.20 \pm 1.50	18.19 \pm 1.04	17.17 \pm 1.18	0.650	0.584

*Significant difference determined by one-way ANOVA

Table 3. Comparison of the mean of iron, ferritin, and UIBC levels between non-pregnant and the various trimesters of pregnant women using the least significant difference (LSD) post hoc.

Parameter	Groups		Mean Difference	P-value
	Non-pregnant state	First trimester		
Iron ($\mu\text{mol/L}$)	12.09 \pm 0.48	13.01 \pm 0.65	-0.93 \pm 0.17	0.270
Ferritin ($\mu\text{g/L}$)	32.93 \pm 3.81	38.50 \pm 5.50	-5.57 \pm 1.69	0.356
Unsaturated iron-binding capacity ($\mu\text{mol/L}$)	65.55 \pm 2.20	59.41 \pm 2.37	6.14 \pm 0.17	0.086
	Non-pregnant status	Second trimester		
Iron ($\mu\text{mol/L}$)	12.09 \pm 0.48	13.92 \pm 0.48	-1.83 \pm 0.00	0.005*
Ferritin ($\mu\text{g/L}$)	32.93 \pm 3.81	24.19 \pm 2.52	8.74 \pm 1.29	0.059
Unsaturated iron-binding capacity ($\mu\text{mol/L}$)	65.55 \pm 2.20	51.86 \pm 1.66	13.69 \pm 0.54	0.000*
	Non-pregnant status	Third trimester		
Iron ($\mu\text{mol/L}$)	12.09 \pm 0.48	13.64 \pm 0.42	-1.56 \pm 0.06	0.021*
Ferritin ($\mu\text{g/L}$)	32.93 \pm 3.81	24.25 \pm 3.29	8.68 \pm 0.52	0.073
Unsaturated iron-binding capacity ($\mu\text{mol/L}$)	65.55 \pm 2.20	53.47 \pm 2.15	12.08 \pm 0.05	0.000*

*Significant difference determined by participant's T-test

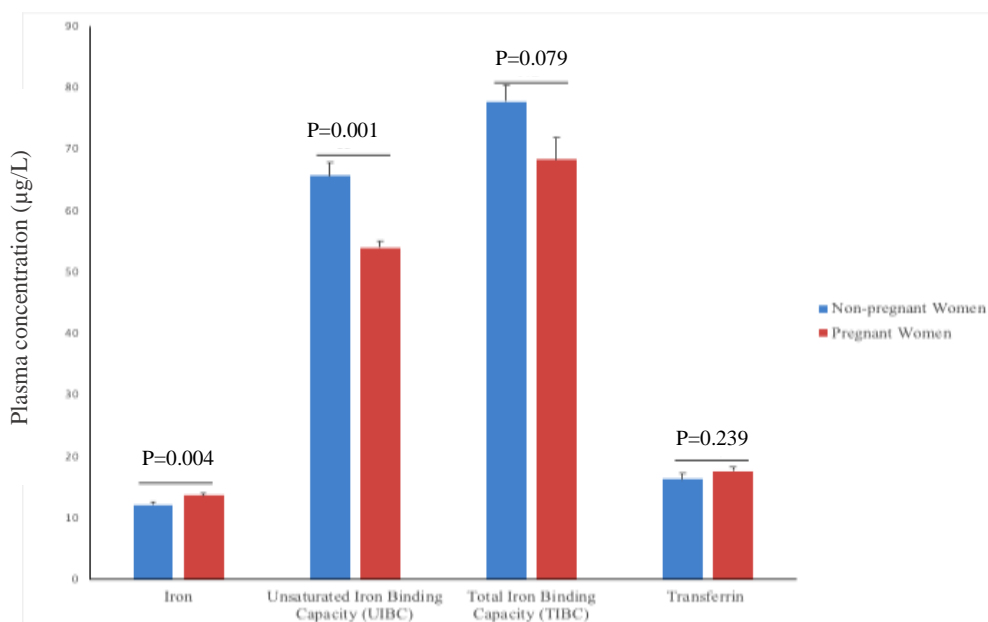


Fig. 1. Comparison of plasma iron, unsaturated iron-binding capacity, total iron-binding capacity and transferrin among non-pregnant and pregnant women. Data are expressed as mean ± SEM.

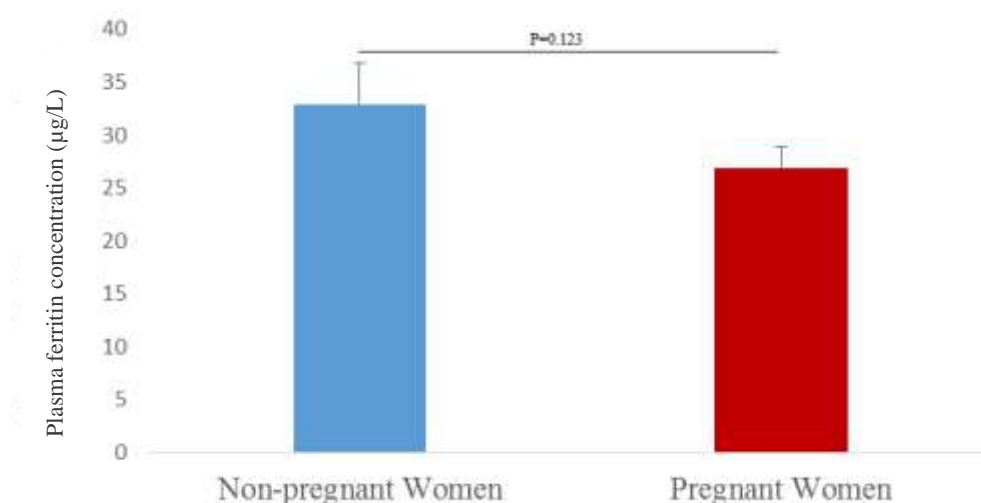


Fig. 2: Comparison of plasma ferritin between non-pregnant and pregnant women. Data are expressed as mean±SEM

Discussion

The development of iron storage deficiency and anemia has been reported in various categories of people. Primarily, ID is widespread, especially among pregnant women in developing countries. Indeed, most cases of non-physiological anemia in pregnancy are due to ID [2].

As a result of iron aggregation, mobilization, or haemo-dilution, iron status seems to decline with

increasing gestational periods [20]. Therefore, several cut-offs values suggestive of iron deficiency across the trimesters of pregnancy have been proposed [21]. Using plasma to evaluate iron homeostasis, this present study evaluated the iron metabolic indices of pregnant women, and take into consideration their various gestational periods. Generally, based on the

gestational age in the assessment and methods of assay and study population, it is quite informative to note that the prevalence of ID in pregnant women differs across the world [22-24].

From this study, ID was more widespread among pregnant women when compared to their non-pregnant counterparts. Although no significant association was observed, the prevalence of ID among pregnant women increases as the gestation age increases. This result is in agreement with similar studies conducted in Singapore, which reported a very high (74%) prevalence of ID among pregnant women at 26-28 weeks than at the early stage of their gestation [24]. It could be attributed to the low intake of iron-rich diets since the most typical form of anemia is nutritional and mostly linked to ID and other dietary elements such as vitamin C, folic acid and vitamin B₁₂. Besides, it could be mentioned that most of these pregnant women had inadequate knowledge about and adherence to good diets recommended for pregnant women. Perhaps, most of these women are among low-income earners who do not have the financial ability to purchase or cook good meals rich in iron. All these could be possible reasons for the ID reported among pregnant women [25, 26, 27]. These studies claimed that knowledge and awareness of ID during pregnancy, in addition to iron supplements, are of utmost significance during antenatal visits [21, 28].

The prevalence of ID reported in this study is higher than 12.3%, 15.7%, and 3.5% prevalence reported by Ejepe [19], Okafor et al. [29] and Erhabor [30] conducted in Cross River, Lagos and the Sokoto States of Nigeria, respectively. This difference in prevalence could be due to the

duration of using iron supplements, which is a useful predictor of reduced risk for ID [19]. Besides, this variance may be attributed to the differences in the geographical location of the study populations.

In the current study, a significant difference was found in the mean iron concentration between pregnant and non-pregnant women. Although it is slightly elevated in pregnant women compared to their non-pregnant counterparts, this change could be attributed to the increase in the demand for iron during the first and second trimesters and possibly it is a result of increased placental and fetal demand [31]. This finding corroborated with the report by Amah-Tariah et al. [32], who recorded significant changes in the plasma iron concentration of pregnant participants. However, they reported a consistent decline in plasma iron levels with an increase in gestational age.

The findings of this study revealed a significant difference in the plasma ferritin across three categories of pregnant women, which indicates that plasma ferritin concentration decreases considerably with the progression of the gestational periods, probably as a result of iron depletion [33, 34]. This observation is supported by a study conducted among pregnant women in Scotland. They reported a high proportion of participants with significantly depleted iron stores associated with increased gestational age, leading to a decline in ferritin concentration [24]. The values of the unsaturated iron-binding capacity from this study showed significant differences between non-pregnant women and their pregnant counterparts. These values declined as the gestational age advanced, which is contrary to the findings of Amah-Tariah et al.,

who reported plasma elevation in UIBC in their participants with increasing gestational age. It could have been a mechanism to ensure a fetal adequate iron delivery based on the decreasing iron concentration with gestation [32]. Furthermore, these values are lower in pregnant women compared to non-pregnant women in the study, and unsaturated iron-binding capacity has been used as a diagnostic indicator of genetic hemochromatosis. Hence, its values remain clinically significant in the management of pregnant women with presumed iron metabolic disorders [35]. Another study also reported a significant increase in UIBC as the age of gestation progressed [36].

Although maternal transferrin production steadily increases during pregnancy [37], which may function to increase iron delivery to the placenta, the plasma transferrin level of pregnant women showed no significant difference compared to those of non-pregnant women. Nevertheless, there was an increased level of transferrin during the second trimester, which is contrary to the findings of Chaudhari et al. [38], who reported an increased level of serum transferrin concentration in the third trimester. The increase in transferrin concentration is likely to be due to elevated steroid levels during pregnancy. Also, a study in Port-Harcourt, Nigeria, reported a consistent increase in plasma transferrin and TIBC with an

increase in gestational age. It suggests a protective mechanism of ensuring proper iron delivery to the developing fetus [32, 39].

Conclusion

This study revealed a high prevalence of ID during pregnancy, which increases with the gestation of pregnant women. It also highlighted the potential importance of routine monitoring and screening for ID at several time points during pregnancy for the timely commencement of ID management.

Iron deficiency has been linked to increased risk of preterm delivery, postpartum bleeding, low birth weight and delayed sensorineural development in infants. Hence, it is essential to periodically evaluate the prevalence of maternal ID and its associated outcomes because it helps to make empirical policies on ID preventive measures and optimization of the outcomes of maternal and fetal wellbeing. Thus, healthcare interventional measures that could address this nutritional and metabolic disorder are recommended.

Conflict of Interest

None declared.

Acknowledgment

None declared.

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