

Original Article

Evaluation of MLL2 (KMT2B) Gene Expression in Colorectal Cancer Tissue Compared with Adjacent Tumor Free Tissue

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Key words

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Background and Aims: An impairment of expression in MLL2 gene involved in histone methylation induces alteration in the methylation patterns at enhancers and causes several cancers including colon cancer. In this study, MLL2 expression was evaluated in North West of Iran.

Materials and Methods: To evaluate the MLL2 expression in patients with colon cancer, 25 tumor samples and 25 samples of tumor margins were collected and analyzed by real-time polymerase chain reaction.

Results: KMT2B level was not associated with age ($p=0.3$), gender ($p=0.05$) and tumor size ($p=0.2$) but had relation with stage of tumor ($p=0.04$).

Conclusions: According to the results obtained from investigating the MLL2 gene expression, it was found that it has a significant relationship with the stage of tumor.

Introduction

Colorectal cancer (CRC) is the fourth leading cause of cancer-related death in Iran [1]. The process of tumor formation is linked with gene mutations [2, 3], deletions or translocations [2, 4]. Apart from genetic harms which are inheritable, the epigenetic changes do play a crucial role in CRC progression [5].

Epigenetic changes are defined as inheritable alterations which occur in the genetic material due to chemical modifications [6] in a way that they do not involve changes in the primary gene nucleotide sequence [7]. These changes may affect chromatin structure and function and alter connection between DNA strands with histone proteins [5, 8]. In eukaryotes, DNA and histone proteins are established into nucleosomes, which, in turn, form the higher-ordered structure of chromatin [9]. The amino-terminal tails of histone proteins are subject to posttranslational modifications [10]. Posttranslational modifications of histone tails regulate chromatin structure and transcription. Epigenetic changes can be divided into several groups, including methylation, ubiquitination, acetylation and the like [11, 12]. Each change can increase or reduce genes expressions, and can therefore affect cellular processes and cause various diseases including cancer [6, 7]. Methylation refers to the transfer of methyl groups on the central histones of nucleosome structure and DNA (without altering the sequence) [10]. Regulation of chromatin accessibility through chromatin remodeling and histone modification (such as methylation and acetylation) is a

critical step in regulating eukaryotic gene transcription [13]. Enzymes that transfer methyl group on lysine or arginine residual in the tail of histones are called histone lysine or arginine methyltransferases [10, 14], lysine may be mono-, di-, or tri-methylated, and arginine residues may be symmetrically or asymmetrically methylated [15]. They comprise a SET domain catalyzing the addition of methyl groups to specific lysine remnants [9, 16]. The mix lineage leukemia (MLL) or KMT2 family exists in multi-protein complexes and is involved in a variety of processes such as normal development and cell growth [6]. Although some of the MLL family members have already been described to be involved in cancer, MLL2 is one of the members of the KMT2 enzyme family whose contribution to cancer is more common than other components of this enzyme family [17]. Histone methylation can result in activation or repression of genes expression. Recent studies have shown evidence that MLL2 (KMT2B) is involved in methylation of the histone H3K4 at enhancers [18], however, H3 lysine 4 methylation (H3K4me) is associated with transcriptional activation and leads to increased expression of genes involved in the transcription [19]. Hence this could be a contributing factor in various cancers including colorectal ones. Several studies conducted on MLL2 gene imply its association with other genes, especially genes involved in cellular processes [20]. KMT2B gene expression has been studied in various cancers the results of which are as follows. In

an investigation conducted at the level of protein and transcription in 2010, Natarajan and his research group found that the level of MLL2 gene increases in the cell line of breast cancer [3, 14]. According to the research carried out by Yin on the situations of this gene on patients with lung cancer, decreased expression of MLL2 gene was resulted [16].

Materials and Methods

Sample collection

Samples were collected from Imam Reza and Shahid Madani Hospitals in a period of one year. The samples were discharged in micro tubes that were first placed Diethyl pyrocarbonate (DEPC) then were transported to liquid nitrogen in the laboratory at the -80 degrees fridge until extraction. Colonoscopy of tumor samples were also transported to the laboratory in the formalin to search for pathological results: For each sample, the pathologic findings, the ethical and protecting confidential patient information were collected from the pathology lab. Patients with CRC as determined by histopathological evaluation were not on any treatment program at that time. RNA extraction from samples had to be performed in an environment free from RNase. Therefore, special care was taken to avoid contamination of utensils and surfaces with RNase enzymes. To deactivate the RNase enzyme, DEPC-treated microtubes were used.

RNA extraction process

Samples of tumor and tumor margin were extracted by using RNA xPlus kit (sigma company), followed by five steps. Step one: putting up 1000 microliter RNA xPlus [21] and

incubating for 5 min; step two: putting up 250 microliter chloroform, centrifuged with 13500 rpm for 15 min, step three: adding 500 microliter Isopropanol, and then incubating for 20 min, and centrifuging with 12000 rpm for 15 min; step four: adding alcohol 75% and centrifuging with 7500 rpm for 8 min; and step five: putting up 15 microliter DEPC water and keeping them in a -80 degrees fridge.

Electrophores gel analysis

In order to check and ensure the extraction process, agarose gel was made. Results in a column with three bands, corresponding to 28s, 18s, 5s were found on the gel. Standard curve of MLL2 with set primers to measure performance of real-time reaction was drawn. The serial dilutions were prepared from cDNA with concentration of (0.1, 0.01, 0.001, and 0.0001).

Real-time polymerase chain reaction (PCR)

PCR test was performed to determine expression of MLL2 in colorectal cancer patients. RNA was DNaseI treated in a 10 µl volume using the takara Kit and then reversely transcribed in the same tubes using combination of oligo (dT) and random hexanucleotide primers. Real-time PCR (10 µl) was performed using the SybrGreen 480 Master Mix that contained 1 µl of 1:5 dilution of complementary DNA (cDNA) and 0.3 µM of each primer. Samples were repeated three times in each well. Temperature cycling conditions were as follows: 1 cycle of 95°C for 10 min., 45 cycles of 95°C for 25 seconds (s), 61°C 30 s, and 72°C 30 s. Mix lineage leukemia (MLL2) and GAPDH (housekeeping gene) primers were respectively as follows:

(forward 5' CGTGGATCCAAGCACCTCCT 3',

reverse 5' TCTTACAGCGCACACAGGCT 3', product: 135 bp); (forward 5' TGTGAACCATGAGAAGTAT 3', reverse 5' CACGATACCAAAGTTGTC 3', product 112 bp). Primer design was done by the Gene runner software. The Ethics Committee of Tabriz University of Medical Sciences approved this study.

Statistical analysis

Statistical analysis of MLL2 expression was performed by unpaired Student's t-test and two-way ANOVA when appropriate. The graphs were generated using SPSS software.

Results

Real-Time analysis result

Statistical analysis indicated that MLL2 expression was not significantly related between normal and tumor samples (Table 1). The linear correlation of MLL2 expression and different factors of age, gender and tumor size ($p=0.3$, $p=0.05$ and $p=0.2$ respectively), revealed no statistical significance. However the stage of tumor showed significant result (Table 2).

Table 1. Comparison of KMT2B expression in different kind of tissue

Kind of tissue	Mean±SD (%)	Fold Change	P-value
Tumoral tissue	0.25±30.50	2.25	0.1
Normal tissue	0.05±31.70		0.1

Table 2. Comparison of KMT2b expression with stage of tumor

Stage	P-value
Stage I	0.40
Stage II	0.71
Stage III	0.04

Discussion

To obtain a global perspective on histone methylation at Lys 4 expression patterns in tumoral and normal cells, Real-time PCR technology was used. Confirming and extending prior conventional studies, it was found that histone methylation at the MLL2 was not related to greater expression of internalizing symptoms except for the stage of tumor. In a research carried out by Prives and Lowe, MLL pathways were associated with GOF p53 oncogenic phenotypes and

therefore lead to cancer progression [22]. The correlation of MLL2 with multiple cellular signaling pathways suggested the potential mechanisms underlying tumorigenesis mediated by MLL2 alterations. These studies are in line with our study supporting the role of mutant MLL2 in driving tumor progression. In contrast, no significant difference was observed between patients with relatively high or low MLL2 expression in tumor samples. Ladopoulos's observations found that the

KMT2B knock down was followed by the decrease of the active chromatin marks and progressive loss of RNA polymerase II binding [23]. Studies by Albert and Helin have shown that expression of KMT2B gene directly connects to transcription factors of specific promoters or enhancer areas [24]. Zhu and coworkers observed that when p53 suffered a missense mutation, it led to an increase in the expression of MLL2 and *moz* genes. However, in the normal state, it fails to reduce the putative expression of these genes [25]. The findings and approaches undertaken here can lay the basis for further understanding of the function of MLL2 and provide

implications for further research on the epigenetic regulator genes that are found to be altered in cancers.

Conclusion

According to the results, investigating the MLL2 gene expression indicated a significant relationship with the stage of tumor.

Conflict of Interest

There are no conflicts of interest or financial involvement with this manuscript as confirmed by all authors.

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References

- [1]. Zhang S, Yuan W, Zhang J, Chen Y, Zheng C, Ma J, et al. Clinicopathological features, surgical treatments, and survival outcomes of patients with small bowel adenocarcinoma. *Medicine (Baltimore)* 2017; 96(31): e7713.
- [2]. Lee BC, Yu CS, Kim J, Lee JL, Kim CW, Yoon YS, et al. Clinico-pathological features and surgical options for synchronous colorectal cancer. *Medicine (Baltimore)* 2017; 96(9): e6224.
- [3]. Natarajan TG, Kallakury BV, Sheehan CE, Bartlett MB, Ganesan N, Preet A, et al. Epigenetic regulator MLL2 shows altered expression in cancer cell lines and tumors from human breast and colon. *Cancer Cell Int.* 2010 30; 10(1): 13.
- [4]. Janke R, Dodson AE, Rine J. Metabolism and epigenetics. *Annu Rev Cell Dev Biol.* 2015; 31: 473-96.
- [5]. Ramachandran S, Henikoff S. Nucleosome dynamics during chromatin remodeling in vivo. *Nucleus* 2016; 7(1): 20-26.
- [6]. Lustberg MB, Ramaswamy B. Epigenetic therapy in breast cancer. *Curr Breast Cancer Rep.* 2011; 3(1): 34-43.
- [7]. Chang CC. Identification of mechanisms and pathways involved in MLL2-mediated tumorigenesis. [Doctoral dissertation], Duke University; 2013.
- [8]. Ata R, Antonescu CN. Integrins and cell metabolism: an intimate relationship impacting cancer. *Int J Mol Sci.* 2017; 18(1): 189.
- [9]. Luger K, Mäder AW, Richmond RK, Sargent DF, Richmond TJ. Crystal structure of the nucleosome core particle at 2.8 Å resolution. *Nature* 1997; 389(6648): 251-60.
- [10]. Bernstein BE, Humphrey EL, Erlich RL, Schneider R, Bouman P, Liu JS, et al. Methylation of histone H3 Lys 4 in coding regions of active genes. *Proc Natl Acad Sci U S A.* 2002; 99(13): 8695-8700.
- [11]. Sanchez J, Arevalo JC. A Review on ubiquitination of neurotrophin receptors: facts and perspectives. *Int J Mol Sci.* 2017; 18(3): 630.
- [12]. Craver LF. Lymphomas and leukemias. *Bulletin of the New York Academy of Medicine.* 1947; 23(2): 79-100.
- [13]. Guo C, Chang CC, Wortham M, Chen LH, Kernagis DN, Qin X, et al. Global identification of MLL2-targeted loci reveals MLL2's role in diverse signaling pathways. *Proc Natl Acad Sci U S A.* 2012; 109(43): 17603-608.
- [14]. Che J, Smith S, Kim YJ, Shim EY, Myung K, Lee SE. Hyper-acetylation of histone h3k56 limits break-induced replication by inhibiting extensive repair synthesis. *PLoS Genetics* 2015; 11(2): e1004990.

- [15]. Martin C, Zhang Y. The diverse functions of histone lysine methylation. *Nature Rev Molr Cell Biol.* 2005; 6(11): 838-49.
- [16]. Yin S, Yang J, Lin B, Deng W, Zhang Y, Yi X, et al. Exome sequencing identifies frequent mutation of MLL2 in non-small cell lung carcinoma from Chinese patients. *Sci Rep.* 2014; 4(1): 6036.
- [17]. Rao RC, Dou Y. Hijacked in cancer: the KMT2 (MLL) family of methyltransferases. *Nature Rev Cancer.* 2015; 15(6): 334-46.
- [18]. Kantidakis T, Saponaro M, Mitter R, Horswell S, Kranz A, Boeing S, et al. Mutation of cancer driver MLL2 results in transcription stress and genome instability. *Gen dev.* 2016; 30(4): 408-20.
- [19]. Kerimoglu C, Agis-Balboa RC, Kranz A, Stilling R, Bahari-Javan S, Benito-Garagorri E, et al. Histone-methyltransferase MLL2 (KMT2B) is required for memory formation in mice. *J Neurosci.* 2013; 33(8): 3452-464.
- [20]. Kawaguchi T, Komatsu S, Ichikawa D, Tsujiura M, Takeshita H, Hirajima S, et al. Circulating micrnas: a next-generation clinical biomarker for digestive system Cancers. *Int J Mol Sci.* 2016; 17(9): 1459.
- [21]. Narayan S, Bader GD, Reimand J. Frequent mutations in acetylation and ubiquitination sites suggest novel driver mechanisms of cancer. *Genome Med.* 2016; 8(1): 55.
- [22]. Prives C, Lowe SW. Cancer: mutant p53 and chromatin regulation. *Nature* 2015; 525(7568): 199-200.
- [23]. Ladopoulos V, Hofemeister H, Hoogenkamp M, Riggs AD, Stewart AF, Bonifer C. The histone methyltransferase KMT2B is required for RNA polymerase II association and protection from DNA methylation at the MagohB CpG island promoter. *Mol Cell Biol.* 2013; 33(7): 1383-393.
- [24]. Albert M, Helin K. Histone methyltransferases in cancer. *Seminars Cell Dev Biol.* 2010; 21(2): 209-20.
- [25]. Zhu J, Sammons MA, Donahue G, Dou Z, Vedadi M, Getlik M, et al. Gain-of-function p53 mutants co-opt chromatin pathways to drive cancer growth. *Nature* 2015; 525(7568): 206-11.