

Original Article

Prevalence of Virulence Factors and Antimicrobial Resistance of *Streptococcus agalactiae* and *Streptococcus uberis* in Ruminant Sub-clinical Mastitic Milk in Iran

Hassan Momtaz^{1*} Ph.D., Mahvash eliyasi¹ M.Sc., Reza Soleimani² DVM, Ali Jazayeri² DVM.

¹Department of Microbiology, Shahrekord Branch, Islamic Azad University, Shahrekord, Iran.

²Faculty of Veterinary Medicine, Shahrekord Branch, Islamic Azad University, Shahrekord, Iran.

ABSTRACT

Article history

Received 4 Oct 2016

Accepted 15 Dec 2016

Available online 15 Mar 2017

Key words

Antibiotic resistance pattern

Mastitic milk

Streptococcus agalactiae

Streptococcus uberis

Virulence genes

Background and Aims: The aim of this study was to determine the virulence and antibiotic resistance properties of *Streptococcus agalactiae* (*S. agalactiae*) and *Streptococcus uberis* (*S. uberis*) isolated from the ruminant's sub-clinical mastitic milk in Iran.

Materials and Methods: Positive samples in the CMT test were cultured and *S. agalactiae* and *S. uberis* have been identified using biochemical tests and polymerase chain reaction (PCR). Virulence and antibiotic resistance genes were studied using PCR. A total of 350 milk samples were collected randomly from 4 major provinces of Iran. We chose 55 bovine, 43 ovine, and 32 caprine herds from these regions.

Results: Totally, 34.52% of bovine, 25% of ovine, and 25% of caprine milk samples were found to be contaminated with *S. agalactiae* and also, 27.38% of bovine, 25% of ovine, and 25% of caprine milk samples were found to be contaminated with *S. uberis*. The most commonly detected virulence genes of *S. agalactiae* were *cfb* (68.75%), *scpB* and *pavA* (43.75%). Also, the most commonly detected pathogenic genes of *S. uberis* were *sua* and *hasB* (83.33%), *cfu* (75.00%) and *skc* (70.83%). Totally, *tetO* (89.28%), *ermA* (82.14%), *mefA* (73.21%) and *tetM* (69.64%) were the most common antibiotic resistance genes in *Streptococcus* isolates. The most resistance were detected against tetracycline (87.5%) and erythromycin (83.92%), respectively.

Conclusions: Our results showed that animal milk samples should be controlled, especially in colder seasons of the year in order to study the presences of *S. agalactiae* and *S. uberis*.

*Corresponding Author: Department of Microbiology, Shahrekord Branch, Islamic Azad University, Shahrekord, Iran. Tel:+983833361045, E-mail: hamomtaz@yahoo.com, hamomtaz@iaushk.ac.ir

Introduction

Mastitis is one of the main health issues in dairy production. Its losses are not only economic, but also issues such as animal health and welfare, milk quality, antibiotic usage and the image of the dairy sector are important reasons to focus on the mastitis control. Accordingly, mastitis is a topic that is well studied worldwide [1]. Mastitis is considered as one of the most costly diseases in dairy herds because the high volume of discarded milk and diminished milk production amount to approximately 80% of financial losses. The remaining 20% of financial losses are attributed to treatment costs, veterinary fees, labor costs, early culling, and death [2]. The major cause of ruminant mastitis is the infection of the udder by pathogenic bacteria among which *Streptococcus agalactiae* (*S. agalactiae*) and *Streptococcus uberis* (*S. uberis*) are the most common pathogens [3, 4]. Several virulence factors, including fibrinogen binding protein (*fnb*), laminin-binding protein (*lmb*), fibronectin-binding protein (*pavA*), β -C protein (*cba*), capsule, C5a peptidase (*scp*), hyaluronate lyase, α -C protein, β -hemolysin/ cytolysin, and CAMP factor (*cfb*) are responsible for *S. agalactiae* infections [5-7]. The *cps* gene cluster coding for the capsule and the *spB* gene coding for surface enzyme ScpB (a C5a peptidase), which causes impairing of neutrophil recruitment and binds fibronectin to promote bacterial invasion of epithelial cells [5]. The C5a peptidase genes were found only in group B streptococci (*scpB* gene) and group A streptococci (*scpA* gene) [6]. The *cba* is a gene codes for alpha-C protein, which is a surface

protein that helps the bacteria to enter the host cells [7]. The *lmb* gene codes for laminine-binding protein (*lmb*), a surface protein that plays a role in the invasion of the damaged epithelium [8]. Presences of fibronectin-binding proteins (*FnbA*, *FnbB* and *FnbC*) have been shown to correspond to the capability of streptococci to adhere to the skin fibroblast [9]. The *S. uberis* virulence factors, including hyaluronic acid capsule, M-like protein, R-like protein, plasminogen activator (*pauA/skc*), and CAMP factor (*cfu*) [10, 11].

Adherence, internalization, and intracellular persistence are important for environmental streptococci like *S. uberis* to establish intramammary infection [11-13]. The *S. uberis* adhesion molecule gene (*sua*), previously referred to as lactoferrin-binding protein was found to play a central role in adherence of *S. uberis* to bovine mammary epithelial cells [14, 15] and thus aid in establishing persistent infection.

The ability to control these infections depends on a detailed knowledge of the epidemiology of the organisms and their environment. Detection of various virulence genes of *S. agalactiae* and *S. uberis* in various resources helps us to know more about the bacterial epidemiology in various parts and resources. However, many reports showed the high presences of antibiotic resistant in these bacteria [16-18]. The high numbers of antibiotic resistance patterns among *S. agalactiae* and *S. uberis* have been coded by of several antibiotic resistance genes [19]. As far as we know, the epidemiology and

prevalence of *S. agalactiae* and *S. uberis* in ruminant mastitic milk is essentially unknown in Iran. Also, veterinarians do not pay attention to proper antibiotic prescription. Therefore, the present study was carried out in order to study the molecular characterization, antibiotic resistance properties and seasonal distribution of *S. agalactiae* and *S. uberis* isolated from ruminant's sub-clinical mastitic milk in Iran.

Materials and Methods

Samples

In order to study the risk factors (origin of milk sample collection and various seasons) for determination the prevalence of *S. agalactiae* and *S. uberis*, from April 2011 to April 2012 (in various seasons of the year), a total of 350 milk

samples were collected randomly from 4 major provinces of Iran (Khuzestan, Isfahan, Fars and Chaharmahal va Bakhtiary). There were 110 bovine, 88 ovine, and 65 caprine herds in these areas and we choose 55 bovine, 43 ovine, and 32 caprine herds from these regions (Table 1). The animals, which their milk samples were collected for this study were clinically healthy and the milk samples showed normal physical characteristics. Samples were collected under sterile hygienic conditions and were immediately transported to the microbiology and biotechnology laboratories of Islamic Azad University, Shahrekord Branch, Iran. The study was approved by Ethics Committee of Islamic Azad university of Shahrekord branch, Iran.

Table 1. Herds size and number of milk samples, which were collected for study the distribution of *S. uberis* and *S. agalactiae* in four provinces of Iran.

Provinces	No. herds in the study			No. herds studied			No. samples per herd			No. milk samples		
	Bovine	Ovine	Caprine	Bovine	Ovine	Caprine	Bovine	Ovine	Caprine	Bovine	Ovine	Caprine
Khuzestan	30	20	25	16	8	13	2-4	2-4	1-2	60	28	20
Isfahan	30	16	10	16	7	6	2-4	2-3	2-3	40	19	14
Fars	30	38	15	15	19	8	2-5	2-3	2-3	50	35	18
Chaharmahal va Bakhtiary	20	14	15	8	9	5	3-5	1-2	3-4	30	18	18
Total	110	88	65	55	43	32	2-5	1-4	1-4	180	100	70

California mastitis test (CMT) and identification of *S. agalactiae* and *S. uberis*

All milk samples were tested using the CMT method [20]. All positive samples were collected through CMT technique was studied for the presences of *S. agalactiae* and *S. uberis*. For isolation of *Streptococcus* species, 0.5 ml of pooled milk sample was enriched in 5 ml of streptococcal selection broth

(HiMedia Laboratories, Mumbai, India) and incubated at 37°C for 6 hrs in 5% CO₂ condition. After enrichment, the samples were streaked onto 5% sheep blood agar (Merck, Darmstadt, Germany) and incubated at 37°C for 24-48 hrs in 5% CO₂ condition. The suspected streptococcal colonies were purified on BHI agar (Merck, Darmstadt, Germany). The purified cultures were

tentatively identified based on Gram's staining and biochemical tests namely, catalase, oxidase, and bile esculin hydrolysis. To differentiate the suspected *S. agalactiae* and *S. uberis* isolated from *Enterococci*, a selective media, KF streptococcal agar (Merck, Darmstadt, Germany) was also used. The colonies were also confirmed using the polymerase chain reaction (PCR) based on the detection of *16SrRNA* gene region of *S. agalactiae* and *S. uberis* described previously [21, 22]. The PCR reaction mixture contained 2.5 µl of 10×Taq polymerase buffer (1.5 mM MgCl₂); 1.0 µl of forward primer (10 µM); 1.0 µl of reverse primer (10 µM); 0.2 µl of dNTP (25 mM), 0.1 µl of Taq polymerase (0.25 u); 5 µl of DNA (50 to 100 ng/µl); add ddH₂O (sterile) to total volume 25 µl. The reaction was carried out in a PCR thermocycler, as follows: 94°C for 4 min.; five cycles of 94°C, 58°C and 72°C for 45 s each step; 20 cycles of 94°C, 58°C, 72°C for 45 s each step; and a step of 72°C for 5 min., at the end of the reaction. Strain of *S. agalactiae* PTCC 1447 (Iranian Research Organization for Science and Technology, Iran) was used as a positive control.

DNA extraction, PCR amplification and gel electrophoresis

Purification of DNA directly from typical colonies was achieved using a DNA extraction kit (DNP™, CinnaGen, Iran) according to the manufacturer's instructions. The concentration of the purified DNA was determined using spectrophotometer (Biophotometer, Eppendorf-Netheler-Hinz GmbH, Hamburg, Germany) and stored at -20°C until further use. Oligonucleotide primers for detection of

virulence factors of *S. agalactiae* and *S. uberis* have been shown in table 2. The *S. agalactiae* isolates were screened for *cfb*, *fnbB*, *pavA*, *lmb*, *scpB*, and *cba* virulence genes, and *S. uberis* isolates were screened for *cfu*, *skc*, *sua*, *lbp*, *hasA*, *hasB*, *hasC*, *gapC* and *oppF* genes. To amplify the virulence genes of *S. agalactiae* and *S. uberis* isolated from ruminant mastitic milk samples, 50 µl of reaction mixture was made containing 20 ng DNA, 1 mM oligonucleotide primers, 0.4 mM of each of dNTPs, 1.5 U Taq polymerase and 1.5 mM MgCl₂. The annealing temperature was varied from 50 to 60°C based on the gene being amplified. The reaction was carried out in a PCR thermocycler, as follows: 94°C for 4 min.; 35 cycles of 94°C, 50-60°C and 72°C for 55 s each step; and a step of 72°C for 8 min., at the end of the reaction. Finally, all PCR products were resolved on 1.5% agarose gel at 90V for 80 min. Gels were stained with ethidium bromide solution (0.5 mg/mL) and photographed under UV light with Uvitech gel documentation (UK).

Antibiotic resistance genes

The macrolide resistance genes *mefA*, *ermA*, and *ermB*, were detected by PCR [23] among all *S. agalactiae* and *S. uberis* isolates. Tetracycline resistance genes *tetM*, *tetO*, *tetT*, *tetW*, *tetL*, *tetQ*, *tetK* and *tetS* were tested by PCR [23-25]. All isolates were tested for the presence of the lincosamide resistance gene *linB* [26]. All isolates were further screened by PCR for the presence of the streptogramin, a resistance gene *vgaB* [27]. All negative results were confirmed in at least three independent assays. Since the positive control sample was not used in regard with detection of some genes, to confirm or

reject the PCR results, the PCR products of the primary positive samples were purified using a PCR product purification kit (Roche Applied Science, Germany) and sent to the MacroGen Co.

(South Korea) for sequencing and sequenced with ABI 3730 XL device and Sanger sequencing method.

Table 2. A set of oligonucleotide primers for potential virulence factors of *S. agalactiae* and *S. uberis*.

Bacteria	Target gene	Primer sequence (5'-3')	Product size (bp)	Annealing temperature (°C)	References
<i>S. agalactiae</i>	<i>Cfb</i>	F-GCTGTTTGAAGTGCTGCTTG R-GACTTCATTGCGTGCCAAC	288	60	23
	<i>fnbB</i>	F-TGATGCTGCAAAAAGAATTGC R-TTACAGCCCCCTTTTGTAGGA	629	53	23
	<i>pavA</i>	F-TTCCCATGATTTCAACAACAAG R-AACCTTTTGACCATGAATTGGTA	495	58	23
	<i>Lmb</i>	F-AGTCAGCAAACCCCAAACAG R-GCTTCCTCACCAGCTAAAACG	397	57	23
	<i>scpB</i>	F-AGTTGCTTCTTACAGCCCAGA R-GGCGCAGACATACTAGTTCCA	567	58	23
	<i>Cba</i>	F-TGCACCGAAGTTACCAGATG R-CAGCCAACCTCTTCGTCGTT	149	56	23
	<i>Cfu</i>	F-TATCCCGATTTGCAGCCTAC R-CCTGGTCAACTTGTGCAACTG	205	59	24
<i>S. uberis</i>	<i>Skc</i>	F-CTCCTCTCCAACAAAGAGG R-GAAGGCCTTCCCCTTTGAAA	800	52	25
	<i>Sua</i>	F-ACGCAAGGTGCTCAAGAGTT R-TGAACAAGCGATTTCGTCAAG	776	58	24
	<i>oppF</i>	F-GGCCTAACCAAAAACGAAACA R-GGCTCTGGAATTGCTGAAAG	419	54	4
	<i>Lbp</i>	F-CGACCCTTCAGATTGGATTC R-TAGCAGCATCACGTTCTTCG	698	53	24
	<i>hasA</i>	F-GAAAGGTCTGATGTGAT R-TCATCCCCTATGCTTACAG	600	58	26
	<i>hasB</i>	F-TCTAGACGCCGATCAAGC R-TGAATTCCYATGCGTCGATC	300	58	26
	<i>hasC</i>	F-TGCTTGGTGACGATTTGATG R-GTCCAATGATAGCAAGGTACAC	300	58	27
	<i>gapC</i>	F-GCTCCTGGTGGAGATGATGT R-GTCACCAGTGTAAGCGTGGA	200	55	24

Antimicrobial resistance pattern

All *S. uberis* and *S. agalactiae* isolates were tested by Kirby–Bauer simple disk diffusion method according to the guidelines from the Clinical and Laboratory Standards Institute [28] for antimicrobial susceptibility tests for bacteria isolated from sub-clinical mastitic milk samples. The following antimicrobials were selected for testing, based on licensing for mastitis treatment in cattle and widespread using

[P10= penicillin (10 u/disk); TE30= tetracycline (30 µg/disk); S10= streptomycin (10 µg/disk); C30=chloramphenicol (30 µg/disk); SXT= sulfamethoxazol (25 µg/disk); GM10= gentamycin (10 µg/disk); ERT10= erythromycin (10 µg/disk), NFX5= enrofloxacin (5 µg/disk); L2= lincomycin (2 µg/disk); CF30= cephalothin (30 µg/disk); CIP5= ciprofloxacin (5 µg/disk); TMP5=trimethoprim (5 µg/disk); F/M300= nitrofurantoin (300 µg/disk); AM10=ampicillin

(10 u/disk)]. Resistance was determined by measurement of inhibition of growth around the antimicrobial disk according to the zone diameter interpretative standards of Clinical and Laboratory Standards Institute [28], and when not available, according to the antimicrobials manufacturers' instructions. *S. uberis* ATCC 27958 and *S. parauberis* ATCC 13386 were used as reference strains.

Statistical analysis

Data were transferred to Microsoft Excel spreadsheet (Microsoft Corp., Redmond, WA, USA) for analysis. Using SPSS 16.0 statistical software (SPSS Inc., Chicago, IL, USA), Chi-square test and Fisher's exact two-tailed test

analysis was performed and the differences were considered significant at values of $p < 0.05$.

Results

Table 3 presents the prevalence of CMT positive milk samples, which were collected from 4 major provinces of Iran. In this study, 84 out of 180 (46.66%), 8 out of 100 (8%), and 4 out of 70 (5.71%) bovine, ovine and caprine milk samples were found to be positive in CMT test. There were significant differences in the prevalence rates of CMT positive milk samples ($p=0.032$). Bovine milk samples were the most common positive CMT samples.

Table 3. Distribution of sub-clinical mastitic milk in ruminants of 4 major province of Iran using the CMT assay

Provinces	No. milk samples			CMT positive (%)		
	Bovine	Ovine	Caprine	Bovine	Ovine	Caprine
Khozastan	60	28	20	28 (46.66)	3 (10.71)	2 (10)
Isfahan	40	19	14	17 (42.5)	2 (10.52)	-
Fars	50	35	18	25 (50)	2 (5.71)	1 (5.55)
Chaharmahal va Bakhtiary	30	18	18	14 (46.66)	1 (5.55)	1 (5.55)
Total	180	100	70	84 (46.66)	8 (8)	4 (5.71)

Table 4 presents the prevalence of *16SrRNA* gene of *S. agalactiae* and *S. uberis* in bovine, ovine, and caprine CMT positive milk samples. Totally, 29 out of 84 bovine (34.52%), 2 out of 8 ovine (25%), and 1 out of 4 caprine (25%) milk samples were found to be contaminated with *S. agalactiae* and also, 23 out of 84 bovine (27.38%), and 1 out of 4 caprine (25%) milk samples were found to be contaminated with *S. uberis*. Totally, the milk samples of Khozestan

and Fars provinces were the most contaminated with *S. agalactiae* and *S. uberis* while the milk samples of Chaharmahal va Bakhtiary province were the less contaminated. There were significant differences for the prevalence of *S. agalactiae* and *S. uberis* ($p=0.021$) and there were significant differences for the prevalence of *S. agalactiae* and *S. uberis* in bovine with ovine and caprine milk samples ($p=0.029$).

Table 4. Detection of the *16SrRNA* genes of *S. uberis* and *S. agalactiae* in CMT positive ruminant milk samples using the PCR technique.

Provinces	CMT positive milk samples			PCR positive (%)					
				<i>S. agalactiae 16SrRNA</i> gene			<i>S. uberis 16SrRNA</i> gene		
	Bovine	Ovine	Caprine	Bovine	Ovine	Caprine	Bovine	Ovine	Caprine
Khozastan	28	3	2	10 (35.71)	1 (33.33)	1 (50)	8 (28.57)	-	1 (50)
Isfahan	17	2	-	6 (35.29)	-	-	4 (23.52)	-	-
Fars	25	2	1	8 (32)	1 (50)	-	7 (28)	-	-
Chaharmahal va Bakhtiary	14	1	1	5 (35.71)	-	-	4 (28.57)	-	-
Total	84	8	4	29 (34.52)	2 (25)	1 (25)	23 (27.38)	-	1 (25)

Table 5 shows the results of PCR assays for detection of genes encoding the virulence factors of *S. agalactiae* and *S. uberis* in bovine, ovine, and caprine milk samples. The PCR product of some these genes are shown in Figure 1 and 2. The most commonly detected genes of *S. agalactiae* were *cfb* (68.75%), *scpB* and *pavA* (43.75%). Also, the most commonly detected genes of *S. uberis* were *sua* (83.33%), *cfu*

(75.00%) and *skc* (70.83%). Bovine milk samples had the highest virulence genes while ovine and caprine milk samples had the lowest virulence genes of *S. agalactiae* and *S. uberis*. There were significant differences ($p=0.036$) for the prevalence of *cfb* gene of *S. agalactiae* with *fnbB* and *lmb* genes and also $p=0.019$ for the prevalence of *sua* and *cfu* of *S. uberis* with *lbp* gene.

Table 5. Distribution of virulence genes of *S. uberis* and *S. agalactiae* in sub-clinical mastitic milk samples.

Species	<i>S. agalactiae</i> virulence genes							<i>S. uberis</i> virulence genes							
	<i>cfb</i>	<i>fnbB</i>	<i>pavA</i>	<i>lmb</i>	<i>scpB</i>	<i>cba</i>	<i>cfu</i>	<i>skc</i>	<i>sua</i>	<i>lbp</i>	<i>hasA</i>	<i>hasB</i>	<i>hasC</i>	<i>gapC</i>	<i>oppF</i>
Bovine	20/29	3/29	14/29	2/29	14/29	4/29	18/23	17/23	19/23	3/23	4/23	19/23	16/23	13/23	15/23
Ovine	1/2	0/2	0/2	0/2	0/2	0/2	0	0	0	0	0	0	0	0	0
Caprine	1/1	0/1	0/1	0/1	0/1	0/1	0/1	0/1	1/1	0/1	0/1	1/1	0/1	0/1	0/1
Total	22/32	3/32	14/32	2/32	14/32	4/41	18/24	17/24	20/24	3/24	4/24	20/24	16/24	13/24	15/24

Table 6 presents distribution of antibiotic resistance genes of *Streptococcal* strains isolated from ruminant sub-clinical mastitic milk samples. The PCR product of some these genes are shown in figure 3. The most common detected antibiotic resistance genes were *tetO* (89.28%), *ermA* (82.14%), *mefA* (73.21%) and *tetM* (69.64%), which were encoding tetracycline and macrolides antibiotics, respectively. Also, *linB* and *vgaB* genes were

detected in six (10.71%) and seven (12.50%) isolates, respectively, which were the less common antibiotic resistance genes. There were significant differences ($p=0.041$) for the prevalence of *tetO* with *tetK*, *tetS*, *linB* and *vgaB* genes and also, $p=0.033$ for the prevalence of *ermA* with *linB* and *vgaB* genes of *Streptococcus* strains. The bovine isolates had the highest amounts of antibiotic resistance genes.

Table 6. Distribution of antibiotic resistance genes of *Streptococcus* strains isolated from ruminant sub-clinical mastitic milk samples.

Species	Antibiotic resistance genes												
	<i>mefA</i>	<i>ermA</i>	<i>ermB</i>	<i>tetM</i>	<i>tetO</i>	<i>tetT</i>	<i>tetW</i>	<i>tetL</i>	<i>tetQ</i>	<i>tetK</i>	<i>tetS</i>	<i>linB</i>	<i>vgaB</i>
Bovine	40/52	44/52	15/52	39/52	46/52	28/52	19/52	15/52	12/52	11/52	11/52	6/52	7/52
Ovine	0/2	1/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2
Caprine	0/2	1/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2
Total	41/56	46/56	15/56	39/56	50/56	28/56	19/56	15/56	12/56	11/56	11/56	6/56	7/56

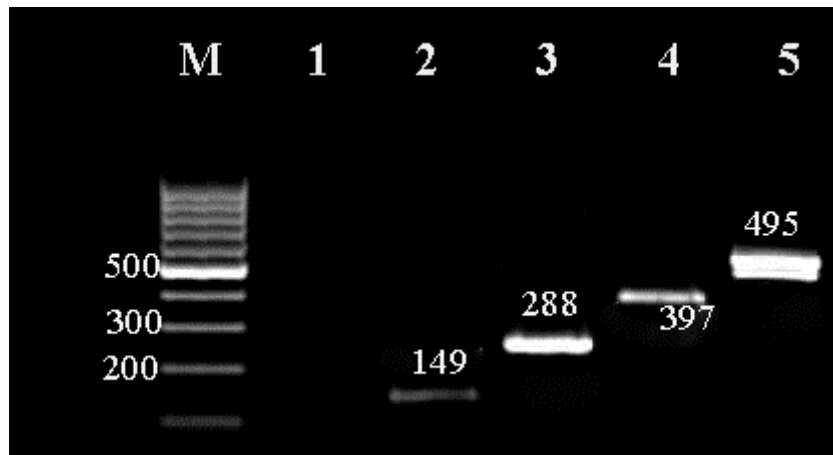


Fig. 1. Agarose gel electrophoresis of PCR products amplified for the *cfb* (288 bp), *pavA* (495 bp), *lmb* (397 bp), and *cba* (149 bp) genes from *S. agalactiae*. M=100bp DNA ladder

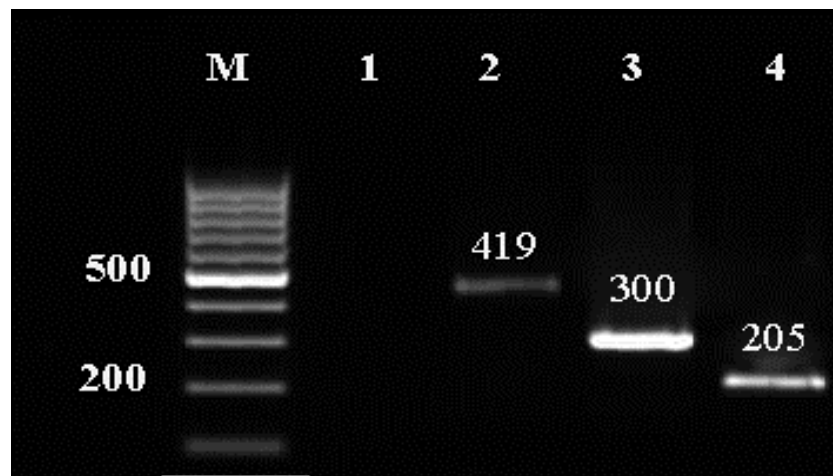


Fig. 2. Agarose gel electrophoresis of PCR products amplified for the *cfu* (205 bp), *oppF* (419 bp), and *hasB* (300 bp) genes from *S. uberis*. M=100bp DNA ladder

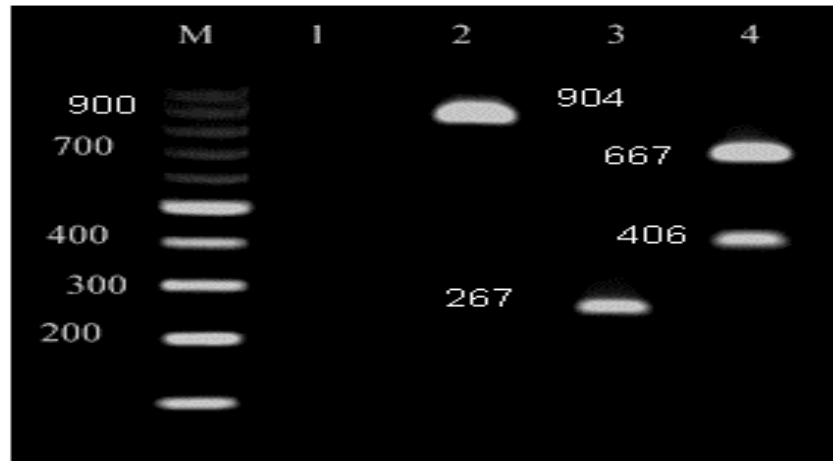


Fig. 3. Agarose gel electrophoresis of PCR products amplified for the *tetL* (267 bp), *tetM* (406 bp), *tetS* (667 bp), and *tetQ* (904 bp) genes from *Streptococcus* strains. M=100bp DNA ladder

Table 7 presents the antibiotic resistance pattern of *Streptococcal* strains isolated from sub-clinical mastitic milk samples. *Streptococcus* strains had the highest antibiotic resistance to tetracycline (87.50%) and erythromycin (83.92%). Lincomycin and streptomycin due to their low antibiotic resistance (5.35% and

7.14%, respectively) are the best drugs for treatment of the cases with *Streptococcus*. Statistical differences were significant between the resistance of *Streptococcus* against tetracycline with lincomycin and streptomycin and erythromycin with lincomycin (p=0.018).

Table 7. Antibiotic resistance pattern of *Streptococcus* strains isolated from ruminant sub-clinical mastitic milk samples (Disk Diffusion Method).

Species	Antibiotic resistance pattern													
	P10	TE30	S10	C30	SXT	GM10	NFX5	L2	CF30	CIP5	TMP5	F/M300	AM10	ERT5
Bovine	14/52	45/52	4/52	18/52	24/52	27/52	26/52	3/52	27/52	24/52	14/52	12/52	24/52	43/52
Ovine	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	2/2
Caprine	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	1/2	2/2
Total	14/56	49/56	4/56	18/56	24/56	27/56	26/56	3/56	27/56	24/56	14/56	12/56	25/56	47/56

P10=penicillin (10 u/disk); TE30=tetracycline (30 µg/disk); S10=streptomycin (10 µg/disk); C30=chloramphenicol (30 µg/disk); SXT=sulfamethoxazol (25 µg/disk); GM10=gentamycin (10 µg/disk); NFX5=enrofloxacin (5 µg/disk); L2=lincomycin (2 µg/disk); CF30=cephalothin (30 µg/disk); CIP5=ciprofloxacin (5 µg/disk); TMP5=trimethoprim (5 µg/disk); F/M300=nitrofurantoin (300 µg/disk); AM10=ampicillin (10 u/disk); ERT5=erythromycin (5 µg/disk)

Table 8 presents the seasonal distribution of *S. uberis* and *S. agalactiae* isolated from ruminant sub-clinical mastitic milk. The milk samples, which were collected in winter season, had the highest contamination rate while the milks,

which were collected in autumn and spring had the lowest contamination rate. Totally, 18.75%, 6.25%, 68.75% and 6.25% of *S. agalactiae* strains were isolated from the milks, which were collected in summer, autumn, winter and spring

seasons and also, 16.66%, 4.16%, 79.16% and 0.0% of *S. uberis* strains were isolated from the milks, which were collected in summer, autumn, winter and spring seasons. There were

significant differences ($p=0.026$) for the rates of contamination between the milk samples, which were collected in winter and spring.

Table 8: Seasonal distribution of *S. uberis* and *S. agalactiae* in ruminant sub-clinical mastitic milk.

Species	No. positive samples		Seasonal distribution							
	<i>S. agalactiae</i>	<i>S. uberis</i>	<i>S. agalactiae</i> (%)				<i>S. uberis</i> (%)			
			Summer	Autumn	Winter	Spring	Summer	Autumn	Winter	Spring
Bovine	29	23	6 (20.68)	2 (6.89)	19 (65.51)	2 (6.89)	4 (17.39)	1 (4.34)	18 (78.26)	-
Ovine	2	-	-	-	2 (100)	-	-	-	-	-
Caprine	1	1	-	-	1 (100)	-	-	-	1 (100)	-
Total	32	24	6 (18.75)	2 (6.25)	22 (68.75)	2 (6.25)	4 (16.66)	1 (4.16)	19 (79.16)	-

Discussion

As far as we know, this present study is the first prevalence report of virulence and antibiotic resistance properties of *S. agalactiae* and *S. uberis* isolated from ruminant sub-clinical mastitic milk in Iran. The results of this present study showed that *S. agalactiae* and *S. uberis* had the high prevalence in sub-clinical mastitic milk of Iran. Totally, *S. agalactiae* was more prevalent than *S. uberis* and it was agreed with previous studies [3, 21]. Our results showed that there were severe seasonal distribution for prevalence of *S. agalactiae* and *S. uberis* in sub-clinical mastitic milk. Totally, these bacteria were more prevalent in the milk samples, which were collected in winter season and it was in agreement with the previous studies [29, 30]. The main reason for the higher prevalence of these bacteria in winter is the fact that during this time climatic events, heat, rain, and thunderstorms, as well as variation of barometric pressure may have influence on the autonomic nervous system. These events

caused reduction in the levels of animal immunity. Therefore, several infections and illnesses have been occurred. Also, animals are more crowded in barn in cold seasons. Therefore, diseases and infections would be disseminated. After analyzing the average temperatures of these 4 provinces in each season (30°C for spring, 45°C for summer, 16°C for autumn and 4°C for winter), it was recognized that the prevalence rate of *S. agalactiae* and *S. uberis* in each season is related with their average temperatures. Our results showed significant differences between relative temperatures of summer with winter. Therefore, the highest levels of animal health care should be performed on the cold months of year. The prevalence rate of infection with *Streptococcus* species between 11 and 47 percent [31]. Totally, 34.52% and 27.38% of bovine milk, 25% and 0.0% of ovine milk, and 25% and 25% of caprine milk samples of our investigation were found to be infected with *S. agalactiae* and *S. uberis*, respectively. The

main source of the infection is the udder of infected ruminants. Although, when hygiene is poor, contamination of the environment may also provide an additional source of infection [21, 31]. These are the most common causes of environmental mastitis in animals. *S. uberis* has been isolated from bedding materials of animals and *S. dysgalactiae* can also be found in the environment of dairy herds [31]. Several investigations have been performed on the prevalence of *S. agalactiae* and *S. uberis* in Iranian dairy herds [3, 32, 33]. Moatamedi et al. [3] showed that *S. agalactiae* and *S. uberis* were causative agents in 20 and 0.83% of CMT positive milk samples, respectively, which was agreed with our results. Ahmadi et al. [32] reported that *S. agalactiae* was isolated from 8% of the 100 samples and 15% were positive in amplification of the *16SrRNA* gene. Our results showed that *cfb* (68.75%), *scpB* and *pavA* (43.75%) were the most common virulence genes of *S. agalactiae* and were *sua* (83.33%), *cfu* (75.00%) and *skc* (70.83%) were the most common virulence genes of *S. uberis* in ruminant milk samples. Beenu et al. [34] reported that out of 27 isolates of *S. agalactiae* none was carrying the *bca* gene, while 6, 7, 8 and 6 isolates were positive for *scpB*, *rib*, *lmb* and *cyiE* genes. All *S. agalactiae* isolates of the Shome et al. [35] study carried the *cfb* gene, only a single isolate possessed the *pavA* gene, while the *cfu* gene was detected in six (85.7 %) *S. uberis* and the *sua* gene was found in four (80 %), which was in line with our results. The *sua* gene of *S. uberis* plays a major role in adherence of bacterium to bovine mammary epithelial cells [14, 15]. The *cba* gene of *S. agalactiae* plays an important role in

bacterial penetration into the host cells [7]. The *lmb* gene plays an important role in the invasion of the damaged epithelium [8]. Therefore, those animals, which infected with streptococcal virulence genes, were more prone to streptococcal mastitis. Because of the importance of antibiotic prescription for the prevention of invasive streptococcal infections especially in mastitis and the concern regarding antibiotic resistance, one of the objectives of this study was to determine the antibiotic resistance properties of *Streptococcus* spp. isolated from ruminant sub-clinical mastitic milk. Our results showed that all of the *Streptococcus* spp., which was isolated from studied milk samples had one or more antibiotic resistance genes. Totally, *tetO* (89.28%), *ermA* (82.14%), *mefA* (73.21%) and *tetM* (69.64%), which were encoding tetracycline and macrolides were the most common detected antibiotic resistance genes. These results are in aligning with the previous studies [19, 36]. In accordance with other reports from all around the world, antibiotic resistance analysis showed that most of the isolates were resistant to tetracycline [37-39]. Also, resistance to fluoroquinolone has been documented in other countries [40]. Erythromycin resistance had been reported from many other sites of the world, too [19, 41, 42]. Streptococcal isolates of our study had the highest antibiotic resistance to tetracycline (87.50%) and erythromycin (83.92%) in disk diffusion method, which were aligned with the previous investigations [19, 43]. Dogan et al. [44] reported that resistance to tetracycline and erythromycin was more common among *S. agalactiae* isolates from humans

than among isolates from cattle. Macrolide and tetracycline resistance among *S. agalactiae* strains have been reported previously from Spain [45]. Resistance against tetracycline, erythromycin, co-trimoxazole, ampicillin and enrofloxacin was 55.5%, 33.3%, 11.1%, 11.1%, and 7.4%, respectively among all mastitic milk borne *S. agalactiae* isolates of previous study [39]. Totally, the presence of antibiotic resistance is different in various geographical regions. Mainly, the high, irregular and unauthorized use of some antibiotics leads to high amounts of antibiotic resistance. It depends on veterinarian's choice of antibiotic for prescription. The high presences of resistance to the studied antibiotic showed the irregular and unauthorized use of it in veterinary treatment in Iran.

Conclusions

Our results showed that animal milk samples should be controlled especially in colder seasons

of the year in order to study the presences of *S. agalactiae* and *S. uberis*. All of the mastitic milk samples harbored *cfb*, *scpB*, *pavA*, *sua*, *hasB* and *cfu* virulence genes and *tetO*, *ermA*, *mefA* and *tetM* antibiotic resistance genes. It is better to use antibiotics in regular and authorized manner based on the results of disk diffusion method. Also, to tetracycline and erythromycin should not be used in cases of Streptococcal infections. It appears that researchers should develop new antibiotics instead of relying on traditional antibiotics for treating animals and even humans with infectious diseases.

Conflict of Interest

All authors declare that there is no conflict of interest.

Acknowledgements

The authors would like to express their deep sense of gratitude and sincere thanks to the staff of the Biotechnology Research Center of Islamic Azad University of Shahrekord Branch in Iran.

References

- [1]. Lam TJGM. Mastitis Control: From Science to Practice, Proceedings of International Conference 30 September-2 October 2008 the Haque, the Netherlands. Wageningen Academic Publishers. 2008.
- [2]. Salwa MH, Ammar MA, Aisha RA; Mona AEI, Hakim AS, Bakry MA, et al. Molecular and virulence characterization of *Escherichia coli* strains isolated from persistent bovine mastitis. J Am Sci. 2011; 7: 614-24.
- [3]. Motamedi H, Seyfiabad Shapouri M, Ghorbanpour M, Jamshidian M, Gouraninezhad S. A polymerase chain reaction based study on the subclinical mastitis caused by *Streptococcus agalactiae*, *S. dysgalactiae* and *S. uberis* in cattle in Ahvaz. Iranian J Vet Res. 2007; 8(3): 260-65.
- [4]. Smith AJ, Kitt AJ, Ward PN, Leigh JA. Isolation and characterization of a mutant strain of *Streptococcus uberis*, which fails to utilize a plasmin derived beta-casein peptide for the acquisition of methionine. J Appl Microbiol. 2002; 93(4): 631-39.
- [5]. Beckmann C, Waggoner JD, Harris TO, Tamura GS, Rubens CE. Identification of novel adhesins from Group B streptococci by use of phage display reveals that C5a peptidase mediates fibronectin binding. Infect Immun. 2002; 70: 2869-876.
- [6]. Franken C, Haase G, Brandt C, Weber-Heynemann J, Martin S, Lämmle C, et al. Horizontal gene transfer and host specificity of beta-haemolytic streptococci: the role of a putative composite transposon containing *scpB* and *lmb*. Mol Microbiol. 2001; 41(4): 925-35.
- [7]. Bolduc GR, Baron MJ, Gravekamp C, Lachenauer CS, Madoff LC. The alpha C protein

- mediates internalization of group B *Streptococcus* within human cervical epithelial cells. *Cell Microbiol.* 2002; 4(11): 751-58.
- [8]. Spellerberg B, Rozdzinski E, Martin S, Weber-Heynemann J, Schnitzler N, Lütticken R, et al. Lmb, a protein with similarities to the LraI adhesin family, mediates attachment of *Streptococcus agalactiae* to human laminin. *Infect Immun.* 1999; 67(2): 871-78.
- [9]. Kline JB, Xu S, Bisno AL, Collins CM. Identification of a fibronectin-binding protein (*GfbA*) in pathogenic group G streptococci. *Infect Immun.* 1996; 64(6): 2122-129.
- [10]. Jiang M, Babiuk LA, Potter AA. Cloning, sequencing and expression of the CAMP factor gene of *Streptococcus uberis*. *Microb Pathog.* 1996; 20(5): 297-307.
- [11]. Oliver SP, Almeida RA, Calvino LF. Virulence factors of *Streptococcus uberis* isolated from cows with mastitis. *Zentralbl Veterinarmed B.* 1998; 45(1-10): 461-71.
- [12]. Tamilselvam B, Almeida RA, Dunlap JR, Oliver SP. *Streptococcus uberis* internalizes and persists in bovine mammary epithelial cells. *Microb Pathog.* 2006; 40(6): 279-85.
- [13]. Patel D, Almeida RA, Dunlap JR, Oliver SP. Bovine lactoferrin serves as a molecular bridge for internalization of *Streptococcus uberis* into bovine mammary epithelial cells. *Vet Microbiol.* 2009; 137(3): 297-301.
- [14]. Fang W, Almeida RA, Oliver SP. Effects of lactoferrin and milk on adherence of *Streptococcus uberis* to bovine mammary epithelial cells. *Am J Vet Res.* 2000; 61(3): 275-279.
- [15]. Chen X, Dego OK, Almeida RA, Fuller TE, Luther DA, Oliver SP. Deletion of sua gene reduces the ability of *Streptococcus uberis* to adhere to and internalize into bovine mammary epithelial cells. *Vet Microbiol.* 2011; 147(3): 426-34.
- [16]. Castor ML, Whitney CG, Como-Sabetti K, Facklam RR, Ferrieri P, Bartkus JM, et al. Antibiotic resistance patterns in invasive group B streptococcal isolates. *Infect Dis Obstet Gynecol.* 2008; 727505.
- [17]. Poyart C, Jardy L, Quesne G, Berche P, Trieu-Cuot P. Genetic basis of antibiotic resistance in *Streptococcus agalactiae* strains isolated in a French hospital. *Antimicrob Agents Chemother.* 2003; 47(2): 794-97.
- [18]. Gherardi G, Imperi M, Baldassarri L, Pataracchia M, Alfarone G, Recchia S, et al. Molecular epidemiology and distribution of serotypes, surface proteins, and antibiotic resistance among group B streptococci in Italy. *J Clin Microbiol.* 2007; 45(9): 2909-916.
- [19]. Usein CR, Grigore L, Georgescu R, Cristea V, Băltoiu M, Străuț M. Molecular characterization of adult-colonizing *Streptococcus agalactiae* from an area-based surveillance study in Romania. *Eur J Clin Microbiol Infect Dis.* 2012; 31(9): 2301-310.
- [20]. Schalm OW, Noorlander DO. Experiments and observations leading to development of the California mastitis test. *J Am Vet Med Assoc.* 1957; 130(5): 199-204.
- [21]. Meiri-Bendek I, Liapkin E, Friedmann A, Leitner G, Saran A, Friedman S, et al. A PCR-based method for the detection of *Streptococcus agalactiae* in milk. *J Dairy Sci.* 2002; 85(7): 1717-723.
- [22]. Bentley RW, Leigh JA. Determination of 16S ribosomal RNA gene copy number in *Streptococcus uberis*, *S. agalactiae*, *S. dysgalactiae* and *S. parauberis*. *FEMS Immunol Med Microbiol.* 1995; 12(1): 1-7.
- [23]. Pires R, Rolo D, Gama-Norton L, Morais A, Lito L, Salgado MJ, et al. Group A Streptococci from carriage and disease in Portugal: evolution of antimicrobial resistance and T antigenic types during 2000-2002. *Microb Drug Resist.* 2005; 11(4): 360-70.
- [24]. Ng LK, Martin I, Alfa M, Mulvey M. Multiplex PCR for the detection of tetracycline resistant genes. *Mol Cell Probes.* 2001; 15(4): 209-15.
- [25]. Aminov RI, Garrigues-Jeanjean N, Mackie RI. Molecular ecology of tetracycline resistance: development and validation of primers for detection of tetracycline resistance genes encoding ribosomal protection proteins. *Appl Environ Microbiol.* 2001; 67(1): 22-32.
- [26]. Bozdogan B, Berrezouga L, Kuo MS, Yurek DA, Farley KA, Stockman BJ, et al. A new resistance gene, *linB*, conferring resistance to lincosamides by nucleotidylation in *Enterococcus faecium* HM1025. *Antimicrob Agents Chemother.* 1999; 43(1): 925-29.
- [27]. Allignet J, El Solh N. Characterization of a new staphylococcal gene, *vgaB*, encoding a putative ABC transporter conferring resistance to streptogramin A and related compounds. *Gene.* 1997; 202(1): 133-38.
- [28]. Clinical and Laboratory Standards Institute (CLSI). Performance Standards for Antimicrobial Disk Susceptibility Tests, Approved standard-Ninth Edition (M2-A9). Clinical and Laboratory Standards Institute, Wayne, PA. 2006.
- [29]. Shathele MS. Weather effect on bacterial mastitis in dairy cows. *Int J Dairy Sci.* 2009; 4(2): 57-66.
- [30]. Olde Riekerink RG, Barkema HW, Stryhn H. The effect of season on somatic cell count and the incidence of clinical mastitis. *J Dairy Sci.* 2007; 90(4): 1704-715.
- [31]. Radostits OM, Gay CC, Hinchcliff KW, Constable PD. *Veterinary Medicine: A textbook of the diseases of cattle, horses, sheep, pigs and goats.* 10th edition, Saunders Ltd., Edingburg. 2007.

- [32]. Ahmadi M, Razavi Rohani SM, Ayremlou N. Evaluation of *Streptococcus agalactiae* detection by PCR in milk and its comparison to other microbiological methods. *Iranian J Microbiol.* 2009; 1(4): 28-31.
- [33]. Soltani M, Pirali E, Shayan P, Eckert B, Rouholahi Sh, Sadr Shirazi N. Development of a reverse line blot hybridization method for detection of some streptococcal/lactococcal species, the causative agents of zoonotic streptococcosis/lactococcosis in farmed fish. *Iranian J Microbiol.* 2012; 4(2): 15-9.
- [34]. Beenu J, Anuj T, Bharat BB, Mayur KJ. Antibiotic resistance and virulence genes in *Streptococcus agalactiae* isolated from cases of bovine subclinical mastitis. *Veterinarski Arhiv.* 2012; 82(5): 423-32.
- [35]. Shome BR, Bhuvana M, Mitra SD, Krithiga N, Shome R, Velu D, et al. Molecular characterization of *Streptococcus agalactiae* and *Streptococcus uberis* isolates from bovine milk. *Trop Anim Health Prod.* 2012; 44(8): 1981-992.
- [36]. Duarte RS, Bellei BC, Miranda OP, Brito MA, Teixeira LM. Distribution of antimicrobial resistance and virulence-related genes among Brazilian group B *streptococci* recovered from bovine and human sources. *Antimicrob Agents Chemother.* 2005; 49(1): 97-103.
- [37]. Garland SM, Cottrill E, Markowski L, Pearce C, Clifford V, Ndisang D, et al. Antimicrobial resistance in group B *streptococcus*: the Australian experience. *J Med Microbiol.* 2011; 60(2): 230-35.
- [38]. Granlund M, Axemo P, Bremme K, Bryngelsson AL, Carlsson Wallin M, Ekström CM, et al. Antimicrobial resistance in colonizing group B *Streptococci* before the implementation of a Swedish intrapartum antibiotic prophylaxis program. *Eur J Clin Microbiol Infect Dis.* 2010; 29(10): 1195-201.
- [39]. Torres RS, Torres RP, Smeesters PR, Palmeiro JK, de Messias-Reason IJ, Dalla-Costa LM. Group a *streptococcus* antibiotic resistance in southern Brazil: a 17-year surveillance study. *Microb Drug Resist.* 2011; 17(2): 313-19.
- [40]. Wehbeh W, Rojas-Diaz R, Li X, Mariano N, Grenner L, Segal-Maurer S, et al. Fluoroquinolone-resistant *Streptococcus agalactiae*: epidemiology and mechanism of resistance. *Antimicrob Agents Chemother.* 2005; 49(6): 2495-497.
- [41]. Domelier AS, van der Mee-Marquet N, Arnault L, Mereghetti L, Lanotte P, Rosenau A, et al. Molecular characterization of erythromycin-resistant *Streptococcus agalactiae* strains. *J Antimicrob Chemother.* 2008; 62(6): 1227-233.
- [42]. Florindo C, Viegas S, Paulino A, Rodrigues E, Gomes JP, Borrego MJ. Molecular characterization and antimicrobial susceptibility profiles in *Streptococcus agalactiae* colonizing strains: association of erythromycin resistance with subtype III-1 genetic clone family. *Clin Microbiol Infect.* 2010; 16(9): 1458-463.
- [43]. Nakamura PA, Schuab RB, Neves FP, Pereira CF, Paula GR, Barros RR. Antimicrobial resistance profiles and genetic characterisation of macrolide resistant isolates of *Streptococcus agalactiae*. *Mem Inst Oswaldo Cruz.* 2011; 106(2): 119-22.
- [44]. Dogan B, Schukken YH, Santisteban C, Boor KJ. Distribution of serotypes and antimicrobial resistance genes among *Streptococcus agalactiae* isolates from bovine and human hosts. *J Clin Microbiol.* 2005; 43(12): 5899-906.
- [45]. Culebras E, Rodriguez-Avial I, Betriu C, Redondo M, Picazo JJ. Macrolide and tetracycline resistance and molecular relationships of clinical strains of *Streptococcus agalactiae*. *Antimicrob Agents Chemother.* 2002; 46(5): 1574-576.